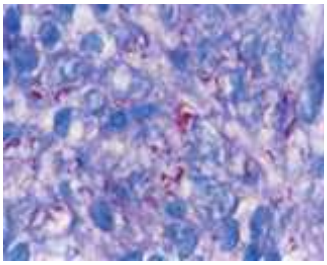
H.S.C. (VOCATIONAL)

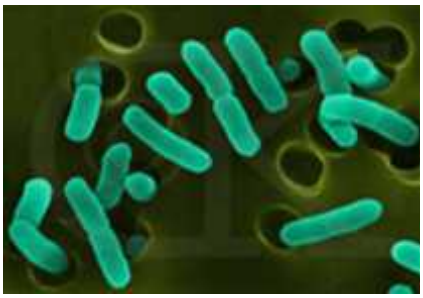
MEDICAL LABORATORY TECHNICIAN

STD: XI (PAPER-3)


MICROBIOLOGY

PRACTICAL HANDBOOK

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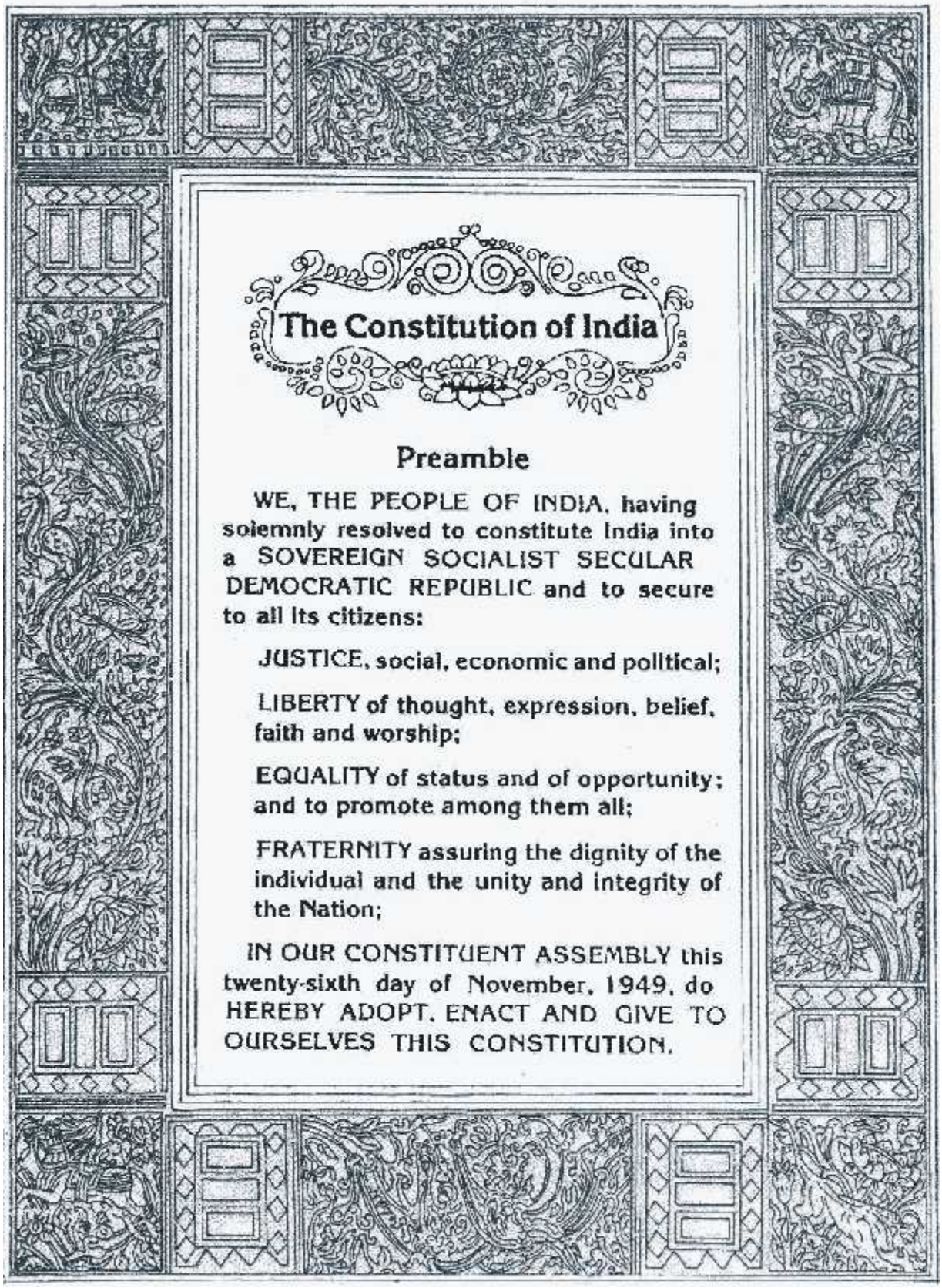

The Constitution of India

Preamble

WE, THE PEOPLE OF INDIA, having solemnly resolved to constitute India into a SOVEREIGN SOCIALIST SECULAR DEMOCRATIC REPUBLIC and to secure to all its citizens:

JUSTICE, social, economic and political;

LIBERTY of thought, expression, belief, faith and worship;

EQUALITY of status and of opportunity; and to promote among them all;

FRATERNITY assuring the dignity of the individual and the unity and integrity of the Nation;

IN OUR CONSTITUENT ASSEMBLY this twenty-sixth day of November, 1949, do HEREBY ADOPT, ENACT AND GIVE TO OURSELVES THIS CONSTITUTION.

****

NATIONAL ANTHEM

Jana-gana-mana-adhināyaka jaya hē
Bhārata-bhāgya-vidhātā
Punjāba-Sindhu-Gujarāta-Marāthā
Drāvida-Utkala-Banga
Vindhya-Himāchala-Yamunā-Gangā
Uchchala-jaladhi-taranga
Tava subha nāmē jāgē, tava subha āsisa māgē,
Gāhē tava jaya-gāthā,
Jana-gana-mangala-dāyaka jaya hē
Bhārata-bhāgya-vidhātā,
Jaya hē, Jaya hē, Jaya hē,
Jaya jaya jaya jaya hē.

****

PLEDGE

India is my country. All Indians are my brothers and sisters.

I love my country and I am proud of its rich and varied heritage. I shall always strive to be worthy of it.

I shall give my parents, teachers and all elders respect and treat everyone with courtesy.

To my country and my people, I pledge my devotion. In their well-being and prosperity alone lies my happiness.

PREFACE

The national policy of education (1986) envisages that the introduction of systematic, well planned and rigorously implemented programme of vocational education is crucial in the proposed educational reorganization. In accordance with the policy of Govt. of India, State govt. of Maharashtra introduces +2 Vocationlization of Education in 1988-89. During last 25 years no substantial efforts has been taken to revamp the curriculum.

Ministry of Human Resource Development, Govt. of India developed the National Skill Qualification Framework (NSQF) to introduce vocational courses according to series of levels of knowledge & skills. Qualifications are made up of vocational standards for specific areas of learning units or units of competency. Units of competency are the specification of the knowledge and skill to the standard of performance expected in the workplace. The unit of competency or National Occupation Standards comprising generic and technical competencies an employee should possess is laid down by the Sector Skill Council of the respective economic or social sector.

The challenges before us were to make smooth transition of curriculum from knowledge based to skill based and rapid technological changes in all sectors of economy. Hence, the few obsolete courses were either merge with core courses or deleted. Hence, in first phase 30 courses were converted into 20 courses. In second phase 20 more courses can be added sector wise as per National Occupational Standards.

I acknowledge the hard team work done by District Vocational Education & Training Officer, who were the coordinators for curriculum designing, theory & practical books writing, along with the vocational teachers of various vocational field & experts from the industry. Shri.S.M.Haste, Joint Director & Shri.A.G.Gavit, Dy.Director has taken the sincere efforts from Directorate to produce the best text material with limited resources & time.

J. D. Bhutange,
Director (Vocational Education)
Directorate of Vocational
Education and Training, Mumbai,
Maharashtra State

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INTRODUCTION TO MEDICAL MICROBIOLOGY

Medical microbiology is the study of microbes that infect humans, the disease they cause, and their diagnosis, prevention and treatment.

The diagnostic microbiology laboratory is engaged in the identification of infectious agents. These infections are broadly classified as viruses, bacteria, mycotic agents and parasites.

Identification of the infectious agent is the principle function of the diagnostic microbiology laboratory. The laboratory also provides information regarding the most effective antimicrobial agent and its dosage to be used for the specific patient.

Microbiology Laboratory Safety and Rules :

- Wear lab coat /apron in the laboratory.
- Do not invite friends into the lab.
- Wear gloves while working in laboratory.
- Wear Shoes in the laboratory.
- Open-toed sandals are not allowed.
- Keep pencils, fingers, etc. out of your mouth, ears, eyes, nose, etc
- Wear face mask where ever required.
- Do not bring food, drink, into the lab.
- Do not take any cultures out of the lab for any reason
- Wipe the bench tops down with disinfectant both before you begin your work and after you have completed your work.
- Keep nonessential books and clothing far away from your work area.
- Wash your hands with soap and water before and after you work in the lab, or if you leave the lab for any reason.
- Mouth pipetting must not be done.
- Any plating/culturing/inoculating must be done in aseptic zone.
- Used glassware, media, etc must be sterilized before and after use.
- Be careful near Bunsen burners.
- Tie long hair back away from the face.
- Turn off burners when not in use.
- Beware of the chemicals and reagents used in biochemical tests.
- Report any accidents (spills, broken glass, cuts, or injuries) to your instructor immediately.
- Dispose of contaminated or used materials quickly and in the manner set forth by your instructor.
- Label all of your tubes and plates exactly as directed by your instructor.
- Place all cultures for incubation exactly where instructed or you risk their loss.
- Any cultures not handled accordingly will be discarded.
- Keep microscopes clean and the objectives free of oil.

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Week No. . .	1st Week
Practical No. . .	1
Title/Aim . .	Operation of instruments
Objectives . .	Student shall be able to operate different insruments used in microbiology laboratory
Requirements . .	Autoclave, pH meter, Hot air oven
Environment . .	MLT laboratory
Procedure. .	**pH METER** **Introduction -**The hydrogen iron conc. or pH is a measure of the acidity or alkalinity of a solution. At given temperature, the product of hydrogen iron conc. and hydroxyl iron conc. is constant. An acid solution has a pH value less than 7 and a basic solution has a pH value greater than 7. A neutral solution has a pH value of 7. **Principle -**When a pair of electrodes is dipped in an aqueous solution, a potential is developed across a thin glass of bulb. The e.m.f. of complete cell formed by joining these two electrodes at a given temperature is E=E (ref) - E (glass) E (ref) = calomel electrode E (glass) = glass electrode **Components-** **Glass electrode -** It consists of a thin bulb blown on to a hard glass tube. It contains 0.1 mole per litre HCL connected to a platinum wire through a silver chloride combination. **Calomel electrode -** It consists of a glass tube containing saturated KCL connected to a platinum wire through mercurous chloride paste. **Operation-** • Turn on and warm up the pH meter. • Immerse the electrodes in a beaker containing standard buffer pH 4. • If the pH reading is not 4, then adjust it to exact 4.0 by using calibration knob. • Wash the electrodes by distilled water and then wipe with soft tissue paper. • Use another standard buffer pH 7 to confirm the standardization. • Now immerse the electrodes in solution under test. • Note the pH readings. **Care and maintainence-** • Glass electrodes must be clean and adequately watered. • For cleaning the glass electrode never use any substance which has absolute alcohol and conc. HCL. • For proper functioning, the calomel electrode must be kept filled with saturated KCL. • When not in use keep dipped in distilled water.

	HOT AIR OVEN - **Use -** Hot air oven is mainly used for (1) Dry sterilization (2) Preparation of anti coagulant bulbs (3) Drying of glass ware. **Principle -** When electricity is passed through the heating coil, the electrical energy is converted to heat energy and the temperature is controlled by a thermostat. **Operation -** • Place the dry articles inside the hot air oven like powdered chemicals. • Drain the washed glass ware first and then arrange inside the oven. • Close the door firmly. Start the electricity. • Control the temperature by using thermostat. • Sterilize for required time. Put off the main switch. • Allow to cool. Remove the articles. **Care and maintenance -** • Put off the main switch when not in use. • Clean the oven after each use. **Autoclave** - Refer to Pratical No. 3.
Clinical significance . .	Knowing operation of various equipments helps to carry out procedures easily and its maintenance and keep instrument in working condition for a longer time.
Skills to be achieved . .	Handling of pH meter, Autoclave and Hot air oven.
Skill evaluation criteria	Use of pH meter - 3marks Preparation of material for sterilization - 4 marks Technique of autoclaving/ Hot air oven - 3 marks Total : 10 marks.
FAQs . .	1. What is principle of hot air oven/autoclave/pH meter ? 2. How care of each equipment is taken ?
Assignment/Activity . .	Study working of each equipment.
Reference . .	Theory topic-3 Handbook of Microbiology

Week No. . .	2nd week
Practical No. . .	2
Title/Aim . .	To disinfect inanimate objects.
Objectives . .	Student shall be able to disinfect inanimate objects.
Principle . .	Disinfection of the work area reduces the chances of contamination of media and cross-contamination of specimens, which help to prevent erroneous results.
Requirements . .	5% phenol solution.
Environment . .	MLT laboratory
Procedure . .	1. Saturate a gauze pad or sponge with the disinfectant (5%phenol) and wipe off the entire work table including edges. 2. Keep a jar of disinfectant in a central position in which all the specimens can be discarded (e.g.throat swab, pus swab etc.) 3. Keep a bucket partially filled with 5% phenol and place under the bench. Discard Specimen bottles in the bucket.
Result . .	The given Inanimate objects are disinfected appropriately.
Clinical significance . .	In order to avoid biological hazard, pathogenic microorganisms handled in the laboratory must be carefully destroyed. The harmless organisms present in the work area might contaminate the media therefore should be removed.
Skills to be achieved. .	Students will disinfect objects properly.
Skill evaluation criteria	Appropriate disinfection of inanimate objects. • At least 10 different glassware - 5 marks. • Selection of disinfectant - 2 marks. • Disinfected edges and corner - 3 marks. Total : 10 marks.
FAQs . .	1. What is disinfection? 2. Name some commonly used disinfectants.
Assignment/Activity . .	• Prepare a list of different disinfectants used. • Disinfect the given specimen.
Reference . .	Theory topic 2 - Handbook of Microbiology

Week No.	. .	3nd week
Practical No.	. .	3
Title/Aim	. .	To do sterilization of glassware, media, etc using autoclave.
Objectives	. .	Student shall be able to do sterilization of glassware, media, etc using autoclave.
Principle	. .	Water boils when its vapor pressure equals that of the surrounding atmosphere. Hence when pressure inside a closed vessel increases, the temperature at which water boils also increases. When steam comes into contact with a cooler surface it condenses to water and gives up its latent heat to that surface.
Requirements	. .	1. Autoclave 2. Articles to be sterilized : Media/glassware/instruments etc. 3. Non – absorbent cotton, paper for wrapping and string.
Environment	. .	MLT laboratory
Procedure	. .	Pre-preparations :- • Test tubes, pipettes, flasks etc glassware must be first clean & dry. • Plug the glassware with non- absorbent cotton. • Liquid contents must not be more than ¾ full & then plugged loosely. • Moist media must be poured in conical flask, after plugging with cotton the necks of the flasks must be covered separately with paper and string. • All the articles to be sterilized must be wrapped with paper & tied with string. *Important* :- Now place the articles in dressing drum with the perforations open of the dressing drum to allow steam to enter. **Sterilization procedure :-** • Fill water in the autoclave through the water – inlet. Open the valve. Water must be filled till the mark indicated. • Close the lid & tighten the screws. • Open air outlet valve. • Plug- in and ON the switch. • Till steam comes out from air outlet valve (saturated steam) • Then close the air – outlet valve. • Now, pressure indicator will move from zero and wait till it comes to required pressure (e.g. 15 lbs). After attaining this pressure count the time (e.g. 20 mins). • Switch OFF the electric supply. Let the pressure drop to zero by opening the outlet valve. • Slowly open the autoclave & remove the dressing drum with articles. Use clean thick cloth duster for handling heated articles & opening the lid.

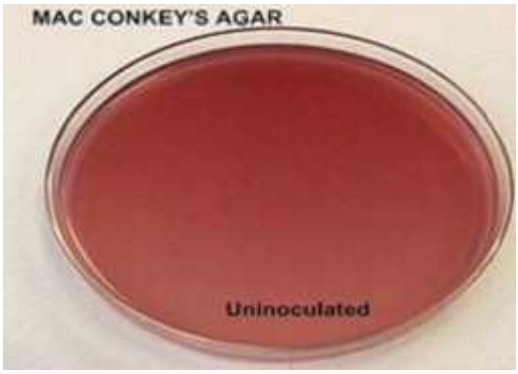
Observations . .	Proper maintenance of pressure, temperature and time during sterilization process.
Result . .	Sterilization of following done • Culture media – 10 lbs pressure for 10 mins. • Glassware, specimen collection bottle – 15 lbs pressure for 20 mins • Infected material -15 lbs pressure for 20 mins.
Clinical significance . .	• Avoids biological hazard. • Sterilization using autoclave is highly effective that kills microorganisms without destroying media.
Skills to be achieved	• Students will operate autoclave and sterilize any given material.
Skill evaluating criteria	1. Proper handling of autoclave. • Correct water level - 1 mark • Required pressure - 1 mark • Required time - 1 mark • Safe removal of glassware - 2 marks. 2. Proper preparation of material for autoclaving. • Correct wrapping of glassware - 2 marks • Plugging of glassware - 2 marks • Labelling - 1 mark. Total : 10 marks.
FAQs	1. What is sterilization ? 2. What are the different methods of sterilization? 3. What is principle of working of autoclave?
Assignment/Activity . .	Sterilize the given material using autoclave.
Reference . .	Theory topic 2 - Handbook of Microbiology

Week No. . .	4th week
Practical No. . .	4
Title/ Aim . .	To sterilize glassware etc using hot air oven.
Objectives . .	Students shall be able to sterilize glassware etc using dry heat (hot air oven).
Principle . .	When electricity is passed through the heating coil, the electrical energy converted to heat energy and the temperature is controlled by a thermostat.
Requirements . .	1. Hot Air Oven 2. Articles to be sterilized : glassware / instruments etc. 3. Non – absorbent cotton, paper for wrapping and string.
Environment . .	MLT laboratory
Procedure . .	1. Place the dry articles inside the hot air oven like powdered chemicals. 2. Drain the washed glass ware first and then arrange inside the oven. 3. Close the door firmly. Start the electricity. 4. Control the temperature by using thermostat. 5. Sterilize for required time. The proper time and temperature for Dry-Heat sterilization is 160 °C (320 °F) for 2 hours or 170 °C (340 °F) for 1 hour. 6. Put off the main switch. 7. Allow to cool. Remove the articles.
Observations . .	Time and temperature maintenance during the process.
Result . .	Given articles are sterilized.
Clinical significance . .	• Avoids biological hazard. • Sterilization using Hot air oven is highly effective that kills microorganisms without destroying media.
Skills to be achieved . .	Students will operate Hot air oven correctly and sterilize any given material..
Skill evaluating criteria	1. Proper handling of Hot air oven. • Correct temperature - 2 marks • Dry glassware - 1 mark • Selection of material - 2 marks 2. Proper preparation of material for sterilization by using Hot air oven. • Correct wrapping of glassware - 2 marks • Plugging of glassware - 2 marks • Labelling - 1 mark. Total : 10 marks.
FAQs . .	1.Explain principle of sterilization by dry heat. 2. What is the temperature for Dry-Heat sterilization?
Assignment/Activity . .	Sterilize the given material using hot air oven.
Reference . .	Theory Topic 2 & 3 - Handbook of Microbiology

Week No.	. .	5th week
Practical No.	. .	5
Title/Aim	. .	To prepare Nutrient Agar.
Objectives	. .	Student shall be able to prepare Nutrient Agar.
Principle	. .	Nutrient agar is used to cultivate differant type of bacteria
Requirements	. .	1. Peptone – 5 Grams 2. Beef extract – 3 Grams 3. Sodium Chloride – 8 Grams 4. Agar – 15 Grams 5. Distilled Water – 1000 ml 6. pH paper
Environment	. .	MLT laboratory
Procedure	. .	1. Place the ingredients except Agar in 1 liter conical flask. 2. Add about 800 ml of distilled water. 3. Dissolve the ingredients either by using the stirrer or if necessary heat. 4. Adjust pH to 7.3 using 1 N NaOH 5. Add Agar and boil for 1-2 min. 6. Make volume to 1000 ml by adding distilled water. 7. Sterilize by Autoclaving at 121°C for 15 min.
Observations	. .	 **Figure 4.1**
Result	. .	Given media is prepared as per the procedure.
Clinical significance	. .	• Used for culturing microorganisms in laboratory to detect the causative organism from given specimen.

Skills to be achieved. .	Students can prepare nutrient agar by using ingredients in required proportion.
Skill evaluating criteria	1. Exact weighing of the ingredients - 4 marks. 2. Media preparation technique - 4 marks 3. Storage - 2 marks. Total : 10 marks.
FAQs . .	• What is the composition of nutrient agar? • What are the different types of media?
Assignment/Activity . .	Prepare 5 nutrient agar plates.
Reference . .	Theory Topic 7 - Handbook of Microbiology

Week No. . .	6th week
Practical No. . .	6
Title/ Aim . .	To prepare Blood Agar.
Objectives . .	Student shall be able to prepare Blood Agar.
Principle . .	It is an Enriched medium used to cultivate different bacteria, mainly hemolytic bacteria
Requirements . .	1. Nutrient Agar- 500 ml 2. Sterile defibrinated blood-25 ml
Environment . .	MLT laboratory
Procedure . .	1. Transfer sterilized Nutrient Agar to 50°C water bath. 2. Add aseptically sterile defibrinated sheep or horse blood. 3. Mix gently. Adjust pH to 7.3.
Observations	 **Figure 5.1**
Result . .	Given media is prepared as per the procedure.
Clinical significance . .	• Used for culturing wide range of pathogens in the laboratory and to detect the causative organism from given specimen. • Differentiate bacteria on the basis of type of hemolysis.
Skills to be achieved. .	Students can prepare blood agar by using ingredients in required proportion.
Skill evaluating criteria	1. Exact weighing of the ingredients - 4 marks. 2. Media preparation technique - 4 marks 3. Storage - 2 marks. Total : 10 marks.
FAQs . .	1. What is the composition of blood agar? 2. How will you differentiate between alpha, beta and gamma hemolysis?
Assignment/Activity . .	Prepare 5 blood agar plates
Reference . .	Theory Topic 7- Handbook of Microbiology

Week No. . .	7th week
Practical No. . .	7
Title/ Aim . .	To prepare MacConkey's Agar.
Objectives . .	Student shall be able to prepare Mac Conkey's Agar.
Principle . .	It is a differential medium used to differentiate between Lactose fermenters and Non lactose fermenters
Requirements . .	1. Peptone - 20 grams 2. Lactose-10 grams 3. Bile salt - 5 grams 4. Sodium chloride - 5 grams 5. Neutral red - 0.075 grams 6. Agar-12 grams 7. Distilled water-1000 ml 8. pH paper
Environment . .	MLT laboratory
Procedure . .	1. Place the ingredients except Agar in 1 liter conical flask. 2. Add about 800 ml of distilled water. 3. Dissolve the ingredients either by using the stirrer or if necessary heat. 4. Adjust pH to 7. 5. Add Agar and heat with frequent agitation and boil for one minute to completely dissolve the medium. 6. Make volume to 1000 ml by adding distilled water. 7. Sterilize by Autoclaving at 121°C for 15 min.
Observations . .	 Figure 7.1
Result . .	Given media is prepared as per the procedure.
Clinical significance . .	Detection of coliform organisms and pathogenic species of enteric bacilli.
Skills to be achieved. .	Students can prepare Mac Conkey's agar by using ingredients in required proportion.
Skill evaluating criteria	1. Exact weighing of the ingredients - 4 marks. 2. Media preparation technique - 4 marks 3. Storage - 2 marks. Total : 10 marks.
FAQs . .	1. What is the composition of Mac conkey's agar? 2. How will you differentiate between lactose fermenters and Non–lactose fermenters?
Assignment/Activity . .	Prepare 5 Mac Conkey agar plates.
Reference . .	Theory Topic 7- Handbook of Microbiology

Week No.	. .	8th week
Practical No.	. .	8
Title/Aim	. .	To study motility of bacteria.
Objectives	. .	The student shall be able to demanstrate of bacteria.
Principle	. .	Under reduced illumination motility is observed at the edge of the drop.
Requirements	. .	1. Cavity slide 2. Cover slip 3. Mineral oil / Liquid paraffin / Vaseline 4. Culture 5. Microscope
Environment	. .	MLT laboratory
Procedure	. .	1. Keep a cover slip on a clean smooth table and put mineral oil/liquid paraffin/Vaseline at the four corners of the cover slip. 2. Put a single loopful of culture in the center of the cover slip. 3. Invert cavity slide (concave side down) over the cover slip and press gently to form a seal. 4. Turn the slide and observe the drop hanging from the cover slip. 5. Observe under low power of microscope and focus the edge of the drop. 6. Now turn to high power & focus with moving the slide. 7. Carefully observe motility of bacteria and report.

Observations

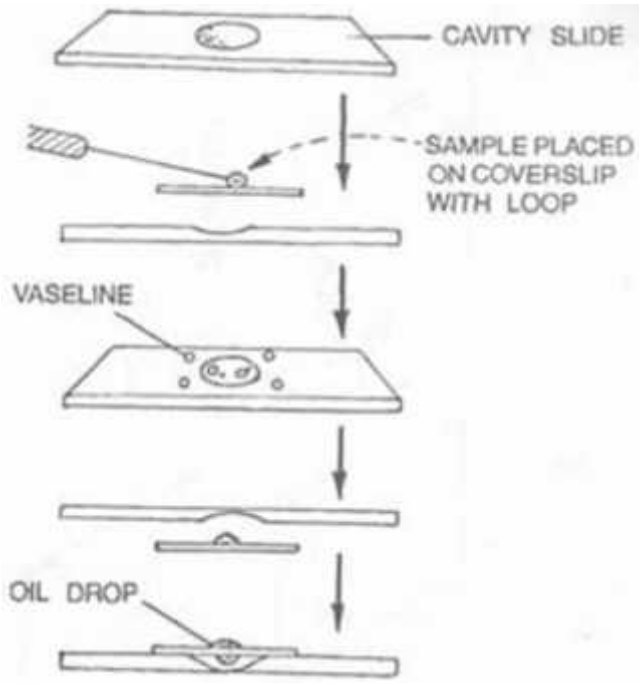


Figure 7.1 Hanging Drop Method

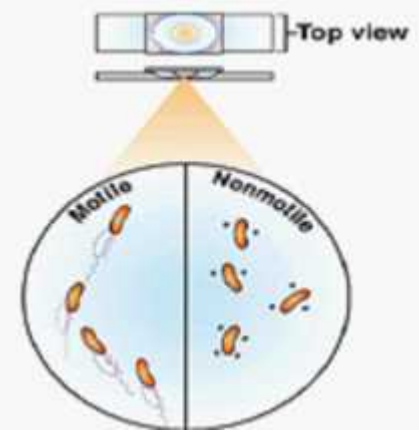


Figure 7.2

Observation :

Morphology	Actively motile	Sluggishly motile	Nonmotile

Result . .	Bacteria are sluggishly motile/actively motile/non-motile.
Clinical significance . .	• Hanging drop technique is used to differentiate between motile and non-motile bacteria. • Motile bacteria possess flagella except spirochetes.
Skills to be achieved . .	Students can observe motility of bacteria by hanging drop method. Students can differentiate between motile and non motile bacteria.
Skill evaluating criteria	• Technique of preparation of hanging drop- 4 marks. • Focusing of the edge of drop under low and high power- 4 marks • Observing the movement of microorganisms- 2 marks Total : 10 marks.
FAQs . .	1. How will you detect that the bacteria are motile or non motile?
Assignment/Activity . .	Perform hanging drop examination of 5 different organisms.
Reference . .	Theory topic 9 – Handbook of Microbiology

Week No. . .	9th week
Practical No. . .	9
Title/Aim . .	Collection of Urine Specimen for bacteriological examination (Patient must not be on antibiotics)
Objectives . .	Student must be able to instruct patient to collect urine specimen for successful isolation of microorganisms.
Principle . .	To obtain midstream sample of the patient taking care that contaminants are avoided and specimen must deliver to the laboratory promptly, as bacteria continue to grow in urine.
Requirements . .	1. Container: Wide-mouthed, screw capped, dry, sterile leak proof bottle provided by the Laboratory. 2. Mid-Stream (clean catch) Sample.
Environment . .	MLT laboratory
Procedure . .	1. First morning discharge of urine is desirable. 2. Ask the patient to wash the genital organ with clean water (do not use soap) before passing the urine. 3. Ask the patient to collect midstream urine sample about (20 ml) after discarding initial portion of urine. 4. After collecting required quantity replace the cap securely. 5. In case of infants a bag is fixed with an adhesive mouth to the genitalia and left for 2-3 hrs. 6. In unavoidable instance catheterization can be done. 7. Clearly label the specimen.
Observations	 **Fig 9.1 Containers for urine**
Result . .	Specimen is obtained taking all the precautions.
Clinical significance . .	Urine specimen is submitted for the diagnosis of urinary tract infection.
Skills to be achieved . .	Students will acquire proper knowledge and significance of sterile containers used for urine collection, its labeling and handling.

Skill evaluating criteria	1. Patient instructions - 2 marks. 2. Knowledge of containers - 2 marks. 3. Selection of containers - 2 marks. 4. Specimen labeling - 1mark. 5. Correct quantity - 1 mark. 6. Specimen handling - 2 marks. Total : 10 marks.
FAQs . .	1. How will you obtain /collect the urine specimen from patient? 2. What instructions you will give the patient before urine sample collection?
Assignment/Activity . .	Collect urine specimen from 5 different patients.
Reference . .	Theory Topic 21 - Handbook of Microbiology

Week No. . .	10th week
Practical No. . .	10
Title/ Aim . .	To collect blood for bacteriological examination.
Objectives . .	Student shall be able to collect blood for bacteriological examination.
Principle . .	Blood is collected by veinipuncture taking all the aseptic precautions during the acute phase of the disease and before any antibiotic administration.
Requirements . .	1. Disposable sterile Syringe & needle (10 ml) 2. Spirit / 70 % alcohol 3. 2% Iodine Solution 4. Tourniquet 5. Sterile swabs 6. Biphasic media
Environment . .	MLT laboratory
Procedure . .	1. Disinfect the skin with spirit at the vein puncture site. 2. Then apply 2% iodine solution wait for minute. 3. Draw blood by venipuncture. 4. Clean with sprit / 70% alcohol 5. Add 5 ml of blood immediately into each collecting bottles before clotting takes place by inserting the needle through the rubber line of the bottle cap.
Result . .	Blood specimen is collected taking all the precautions for bacteriological examination.
Clinical significance . .	Blood is probably the most important specimen submitted to the bacteriological laboratory for culture for the diagnosis of septicemia, bacteraemia, PUO, etc.
Observations.-	The method of collecting blood is illustrated by image as follows :—

Figure 10.1

Skills to be achieved ·	Collection of blood by vein punctures under aseptic precautions.

Skill evaluating criteria	1. Patient instructions - 2 marks. 2. Knowledge of containers - 2 marks. 3. Selection of containers - 2 marks. 4. Specimen labeling - 1mark. 5. Correct quantity - 1 mark. 6. Specimen handling - 2 marks Total : 10 marks.
FAQs . .	1. How will you collect blood from patient? 2. What instructions you will give the patient before blood collection?
Assignment/Activity . .	Collect blood specimen from 5 different patients.
Reference . .	Theory Topic 21 - Handbook of Microbiology

Week No. . .	11th week
Practical No. . .	11
Title/Aim . .	To know the technique of collecting stool sample for bacteriological study.
Objectives . .	Students must know the technique of collecting stool sample for bacteriological study.
Requirements . .	1. Containers- Screw capped, clean, dry, wide mouth, 250 ml bottle or bed pan or sterile swab in case of rectal swab. 2. Cary–Blair medium in case of delay/transport of Specimen.
Environment . .	MLT laboratory
Procedure . .	1. In case of freshly discharged faeces give the container and culture within 2 hrs. 2. In case of bed-pan specimen transfer small portion of specimen to a clean dry container. 3. For rectal swabs- Take the swab after ejection of stool. Insert the swab well, swab the rectal wall. Take 2 swabs. 4. Transportation: Transfera portion of Specimen in to Cary –Blair transport medium. 5. Refrigerate the Specimen if prolonged delay. 6. Label the Specimen correctly.
Observations   **Figure 11.1 Containers for Stool Collection.**	
Result . .	Specimen is obtained taking all the precautions
Clinical significance . .	To see for the presence of an enteric pathogen causing acute intestinal infection (diarrhoea).
Skills to be achieved . .	Proper technique of collection of specimen. Knowledge of containers.
Skill evaluating criteria	1. Patient instructions - 2 marks. 2. Knowledge of containers - 2 marks. 3. Selection of containers - 2 marks. 4. Specimen labeling - 1 mark. 5. Correct quantity - 1 mark. 6. Specimen handling - 2 marks. Total : 10 marks.
FAQs . .	1. How will you obtain /collect the stool specimen from patient? 2. Which are the different containers for stool specimen?
Assignment/Activity . .	Collect stool specimen from 10 different patients
Reference . .	Ref. Theory Topic 21- Handbook of Microbiology

Week No. . .	12th week
Practical No. . .	12
Title/ Aim . .	Collection of Throat Swab and Vaginal Swab
Objectives . .	Students must be aware of appropriate Specimen collection for identification of causative organism.
Principle . .	Obtain specimen without any contaminants
Requirements . .	Sterile cotton swabs within test tube.
Environment . .	MLT laboratory
Procedure . .	**Throat Swab :** • Tilt the head of the patient slightly backwards. • Ask him to open his mouth wide. • Press the tongue with the tongue depressor using left hand. • With right hand pass the swab well over both the tonsils and area of inflammation and return to container. Take 2 Swabs **Vaginal Swab :** • It is taken by a physician/trained nurse or in some cases by an experienced technician. • Collect the vaginal discharge with sterile cotton swab. • Put it into Amies Transport Medium under sterile condition. • Make smears for gram staining. • The Specimen should not be refrigerated.

Observations

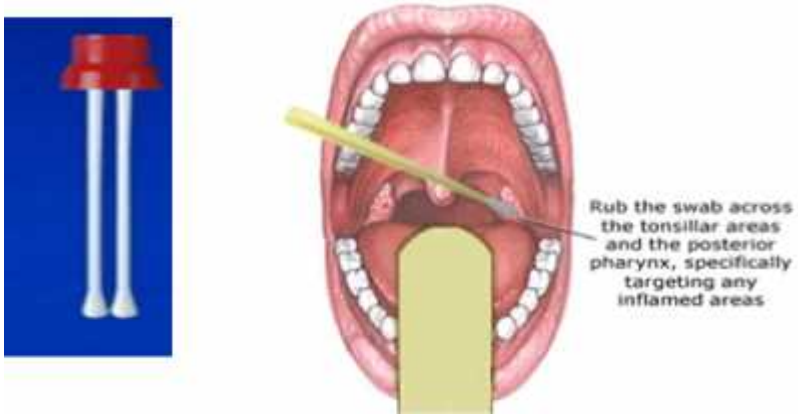


Figure 12.1 Collection of Throat Swab

Result . .	All the specimen collection technique must contain only those organisms from which it is collected avoiding contamination. It should be appropriate for further identification of causative organism
Clinical significance . .	**Throat swab :** Diagnosis of streptococcal sore throat which might lead to serious problems, such as rheumatic fever, scarlet fever and acute glomerulonephritis. **Nasopharyngeal swab :** Diagnose whooping cough and diphtheria. **Vaginal swab :** Diagnosis of Gonorrhoea

Skills to be achieved . .	Collection of throat swab and vaginal swab
Skill evaluating criteria	1. Patient instructions - 2 marks. 2. Knowledge of containers - 2 marks. 3. Selection of containers - 2 marks. 4. Specimen labeling - 1 mark. 5. Correct quantity - 1 mark. 6. Specimen handling - 2 marks. Total : 10 marks.
FAQs . .	1. How will you obtain/collect Throat Swab from patient ? 2. How will you collect vaginal swab from a patient ?
Assignment/Activity . .	Take 5 specimens of throat swab
Reference . .	Ref. Theory Topic 21 - Handbook of Microbiology

Week No.	. .	13th week
Practical No.	. .	13
Title/ Aim	. .	To do Gram staining of given specimen.
Objectives	. .	Student shall be able to identify gram positive and gram negative organisms after staining with gram staining technique.
Principle	. .	Bacteria pick up purple color when stained with crystal violet. Iodine fixes the stain to the bacterial cell. Acetone- alcohol decolorizes gram negative bacteria whereas gram positive bacteria retain the purple color counterstaining with saffranine or basic fuschin stain. Gram negative bacteria appear red.
Requirements	. .	• Crystal violet • Gram's Iodine • Decolourizer (alcohol-acetone) • Saffranine solution • Bunsen burner and Nichrome wire loop • Specimen
Environment	. .	MLT laboratory
Procedure	. .	1. Make a thin smear of the specimen. Allow it to dry. 2. Pass the slide two to three times through flame until it feels comfortably warm on the back of the hand (Heat fixing) 3. Place the slide on staining rack. Add Crystal violet stain for one minute. Wash gently with water 4. Add Gram's Iodine for one minute. Drain the Iodine. Do not wash. 5. Pour Acetone-alcohol on smear till no more blue color comes out from smear (Decolourization) 6. Wash with water. 7. Stain with Saffranine for 10 seconds (Counterstaining) 8. Wash with water. Dry. 9. First observe under low power objective (10x), then under high power (45x) and finally under oil immersion lens using a drop of Cedar wood oil

Observations

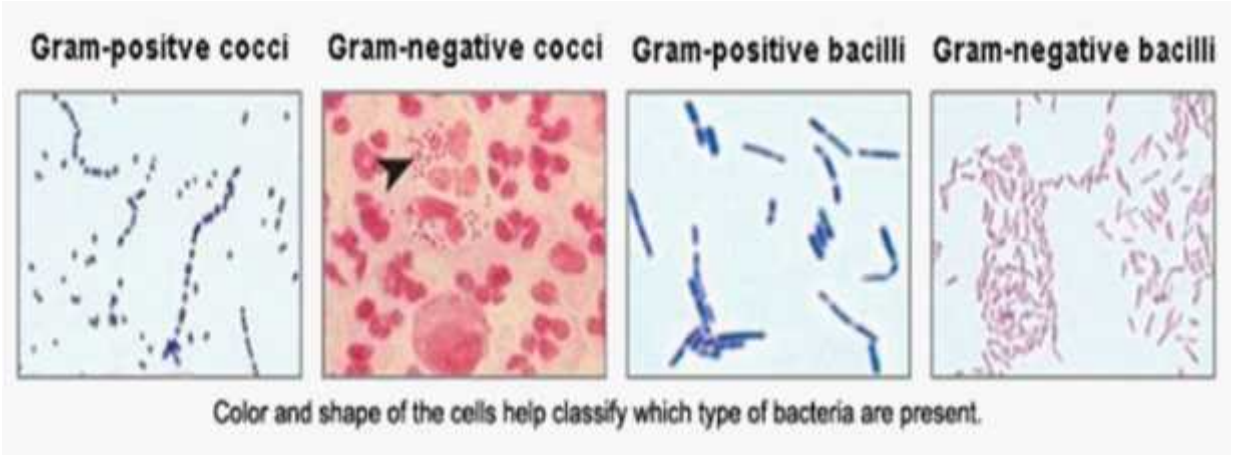


Figure 13.1 Gram Staining

Result . .	**Gram positive** bacteria appear purple and **Gram negative** bacteria appear pink in color.
Clinical significance	• Differentiating into Gram positive and Gram negative is helpful in determining the subsequent biochemical tests and media for their culture. • A preliminary report can be submitted which might help the physician to initiate therapy well before culture results are available
Skills to be achieved . .	Proper staining of specimen with Gram's technique in order to differentiate Gram positive and Gram negative bacteria.
Skill evaluating criteria	1.Heat fixation of smears - 2 marks. 2. Working under aseptic conditions - 2 marks. 3. Selection of stains - 1 mark. 4. Staining technique - 5 marks. Total : 10 marks.
FAQs . .	1. How you will do gram staining of given specimen? 2. Report the findings of gram staining.
Assignment/Activity . .	Do gram staining of urine sample, throat swab, nasal swab, sputum specimen (2 samples each)
Reference . .	Theory Topic 5 - Handbook of Microbiology

Week No. . .	14th week
Practical No. . .	14
Title/ Aim . .	To do Albert's staining
Objectives . .	Student shall be able to identify Metachromatic granules of Corynebacterium diphtheriae after staining with Albert's stain.
Principle . .	Malachite green stains the metachromatic granules bluish black and the bacterial body green.
Requirements . .	• Albert's solution A • Albert's solution B • Bunsen burner • Nichrome wire loop • Specimen
Environment . .	MLT laboratory
Procedure . .	• Make a thin smear of specimen. Allow to dry. • Pass the slide two to three times through flame until it feels comfortably warm on the back of the hand (heat fixing)· • Place the slide on staining rack. Flood with Albert's solution A for five minutes. • Drain the solution. Do not wash with water. • Flood with solution B and keep for one to two minutes. • Wash gently under running tap water. • Drain. Dry. And examine under oil immersion objective (100x).

Observations

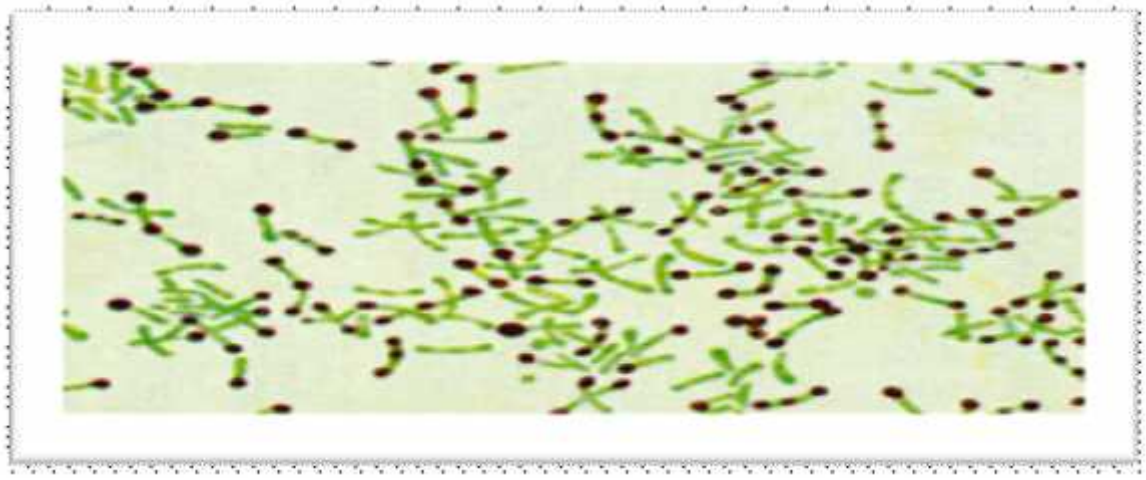


Figure 14.1 ALBERT STAIN - C. Diphtheriae

Result . .	Metachromatic granules appear bluish black, Bacilli green or bluish green.
Clinical significance . .	To detect Corynebacterium diphtheria

Skills to be achieved . .	Skill to do albert staining and keenly observe the metachromatic granules and identify the granules.
Skill evaluating criteria	1. Heat fixation of smears - 2 marks. 2. Working under aseptic conditions - 2 marks. 3. Selection of stains - 1 mark. 4. Staining technique - 5 marks. Total : 10 marks.
FAQs	1. How you will do Albert's staining ? 2. Report the findings of Albert's staining.
Assignment/Activity . .	Do Albert's staining of 3-5 specimen.
Reference	Theory Topic 5 - Handbook of Microbiology.

Week No. . .	15th week
Practical No. . .	15
Title/ Aim . .	To do Z.N. staining (Ziehl-Neelsen Staining)
Objectives . .	Students shall be able to identify Acid Fast organisms after staining with Z.N. stain
Principle . .	Mycobacteria are extremely difficult to stain by ordinary methods because of the lipid containing cell walls called mycolic acid. Heat is applied in hot stain method for detection of Mycobacterium tuberculosis and cold stain method is used for detection of Mycobacterium leprae. Acid Fast bacilli resist de-staining with strong decolorizing agent such as alcohol and strong acids. Acid Fast negative are counterstained with methylene blue stain.
Requirements . .	• Carbol fuchsin stain • 20% Sulphuric acid (for M. Tuberculosis) or 5% Sulphuric acid (for M. leprae) • Methylene blue stain • Bunsen burner • Nichrome wire loop • Specimen
Environment . .	MLT laboratory
Procedure . .	1. Make a thin smear of the specimen. Allow it to dry. 2. Pass the slide two to three times through flame until it feels comfortably warm on the back of the hand (heat fixing). 3. Place the slide on staining rack. Add Carbolfuchsin stain. 4. Heat gently with flame until steam rises. Avoid boiling and continue heating for about five minutes. Do not allow stain to dry. 5. Wash the slide with water. 6. Cover the slide with 20% Sulphuric acid for one minute. (Decolourization) 7. Wash the slide with water. 8. Cover the slide with methylene blue for one minute (Counter staining) 9. Wash with tap water. Allow the water to drain off completely. Dry. 10. Observe the slide first under low power objective (10x) and then under oil immersion objective (100x).
Observations • Acid Fast organisms are bright red bacilli on blue background. • Other organisams and cells appear blue.	

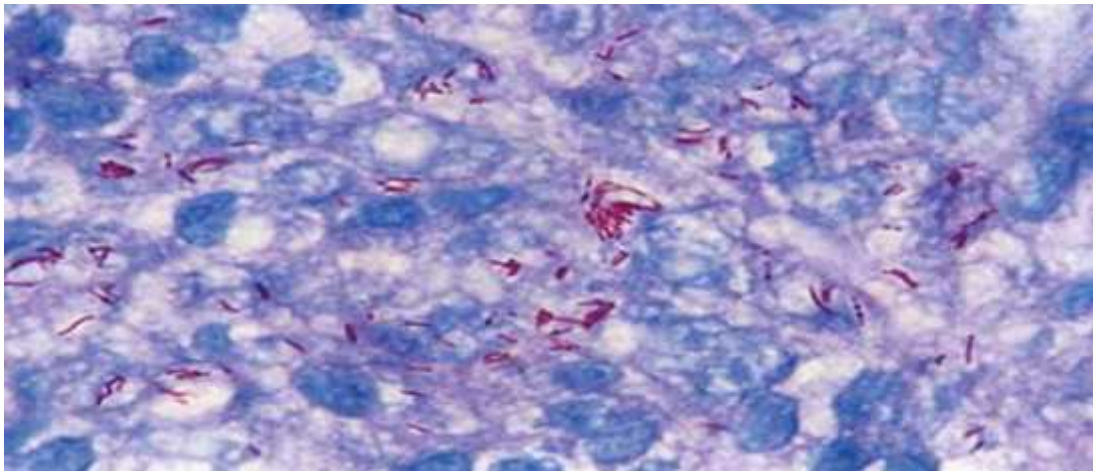


Figure 15.1 Z.N. Staining

Result

No. of Acid Fast Bacteria in 10 fields	Report
None	Absent
2 to 2	Positive, +
10 to 10	Positive, ++
10 to 100	Positive, +++
Above 100	Positive, ++++

Clinical significance . .	To detect Mycobacterium tuberculosis
Skills achieved . .	Skillfully do acid fast staining without any error.
Skill evaluation criteria	1. Heat fixation of smears - 2 marks. 2. Working under aseptic conditions - 2 marks. 3. Selection of stains - 1 mark. 4. Staining technique - 5 marks. Total : 10 marks.
FAQs . .	How you will do acid fast staining of given specimen?
Assignment/Activity . .	Do Acid fast staining of 5 Sputum specimen.
Reference . .	Theory Topic 5 - Handbook of Microbiology

Week No. . .	16th week
Practical No. . .	16
Title/ Aim . .	To do Streak culture method.
Objectives . .	Learning technique of streak culture to enable the students to isolate microorganisms (bacteria).
Principle . .	Streaking is done on primary plate to provide information on the number of types of bacteria in the sample and pure culture is obtained from secondary plate.
Requirements . .	• Petri plates with media. • Specimen / Culture.· • Inoculating loop. • 2 Burners / Safety hood (Aseptic zone). • Marker pen.
Environment . .	MLT laboratory
Procedure . .	Apply this method only for specimens with a low number of organisms.
	Continuous method **Procedure:** 1. Swab the table, burners with a disinfectant. Ignite two burners. All the plating / streaking technique must be done in- between 2 burners is aseptic zone / safety hood. 2. At the back of the plate mark a straight line exactly in the center (diameter). 3. Open the plate with your left hand, thumb & index finger on the lid & other three fingers at the base of the plate. 4. Place a loopful of the specimen / inoculums on the marked central line of the plate touching the media and spread in a single line across the diameter of the plate. 5. Streak back & forth (zigzag) in lines perpendicular to the initial streak, covering the entire plate. 6. Now, turn the plate 90° and streak back and forth in lines parallel to the initial streak, again covering the entire plate.

Observations

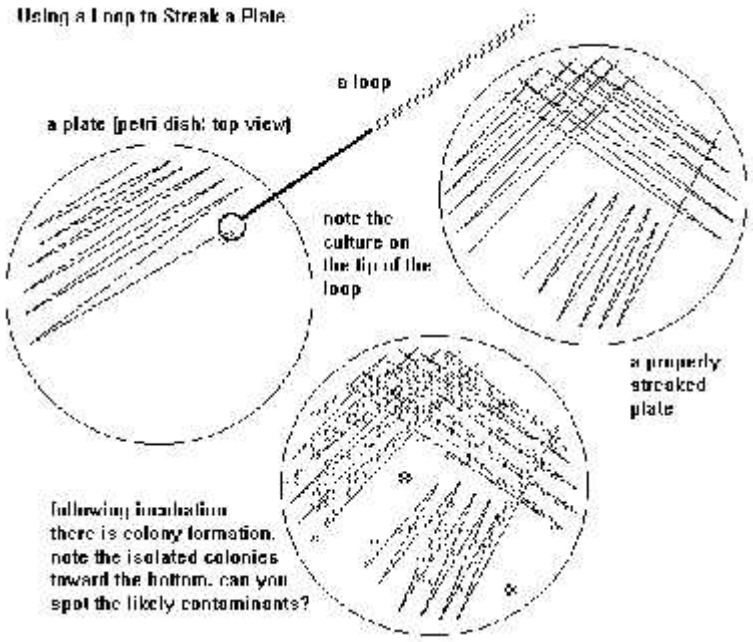


Figure 16.1 Streak culture - Continuous Method

Result . .	Well isolated colonies obtained.
Clinical significance . .	Identification of infectious agent.
Skills achieved . .	1. Use of inoculating loop - 3 marks 2. Technique of aseptic transfer - 3 marks 3. Actual streaking - 3 marks. 4. Obtaining isolated colonies - 1 mark. Total : 10 marks.
FAQs . .	How you will do streaking on primary culture plate?
Assignment/Activity . .	Do streak culture - continuous method of few specimen.
Reference . .	Theory Topic 8 - Handbook of Microbiology

Week No.	. .	17^{th} week
Practical No.	. .	17
Title/ Aim	. .	Identification of isolated bacteria by biochemical test-TSI (triple sugar iron) test
Objectives	. .	The student shall be able to do TSI test.
Principle	. .	• It indicates whether a bacterium ferments glucose only, lactose and sucrose also, with or without gas formation, besides indicating H_2S production as well. • The medium is distributed in tubes, with a butt and slant. After inoculation if slant remains red and the butt becomes yellow,all the sugars - glucose,lactose and sucrose –are fermented. Bubbles in butt indicate gas production and blackening shows H_2S formation.
Requirements	. .	• TSI slant • Incubator • Culture of organism • Inoculating loop and straight needle
Environment	. .	MLT laboratory
Procedure	. .	Streak the TSI slant with a loop and stab with a straight needle. Incubate at 37^0C for 18 to 24 hrs.

Observation

• Alkaline slant (red) and acid butt (yellow) with or without gas production (breaks in the agar butt) :

Only glucose fermentation has occurred. The organisms preferentially degrade glucose first. Since this substrate is present in minimal concentration, the small amount of the acid produced on the slant surface is oxidized rapidly. The peptones in the medium are also used in the production of alkali. At the butt, the acid reaction is maintained because of the reduced oxygen tension and slower growth of the organisms.

• Acid slant (yellow) and acid butt (yellow) with or without gas production :

Lactose or sucrose fermentation has occurred. Since these substances are present in higher concentrations, they serve as substrates for continued fermentative activities with maintenance of an acid reaction in both the slant and the butt.

• Alkaline slant (red) and alkaline butt (red) or no change (orange-red) butt :

No carbohydrate fermentation has occurred. Instead; peptones are catabolized under anaerobic and /or aerobic conditions resulting in alkaline pH due to production of ammonia. If only aerobic degradation of peptones occurs, the alkaline reaction is evidenced only on the slant surface. If there is aerobic and anaerobic utilization of peptone, the alkaline reaction is present on the slant and the butt.

• Hydrogen sulfide (H_2S) production :

Some bacteria utilize thiosulfate anion as a terminal electron acceptor, reducing it to sulfide. If this occurs, the newly-formed hydrogen sulfide (H_2S) reacts with ferrous sulfate in the medium to form ferrous sulfide, which is visible as a black precipitate. The blackening of the medium is almost always observed in the butt (bottom) of the medium.

• Carbon dioxide (CO_2) production :

It is recognized simply as bubbles of gas between the agar and the wall of the tube or within the agar itself. The carbon dioxide production is sufficient to split the agar into two or more sections. To obtain accurate results, it is absolutely essential to observe the cultures within 18-24 hours following incubation. This will ensure that the carbohydrate substrates have not been depleted and that degradation of peptones yielding alkaline end products has not taken place.

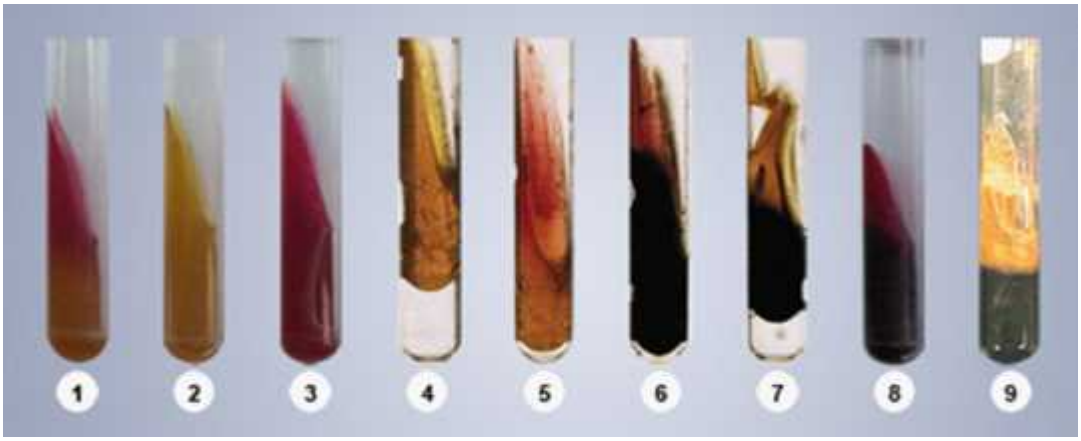


Figure 17.1 TSI test

Result

Sr. No.	Result(Slant/Butt)	Symbol	Interpretation
1	Red/Yellow	K/A	Glucose fermentation only, peptone catabolized.
2	Yellow/Yellow	A/A	Glucose and lactose and/or sucrose fermentation.
3	Red/Red	K/K	No fermentation, Peptone catabolized.
4	Yellow/Yellow with	A/A,G	Glucose and lactose and/or sucrose bubbles.fermentation, Gas produced.
5	Red/Yellow with bubbles.	K/A,G	Glucose fermentation only, Gas produced.
6	Red/Yellow with bubbles and black precipitate.	K/A,G,H_2S	Glucose fermentation only, Gas produced, H_2S produced.
7	Yellow/Yellow with sucrose bubbles and black precipitate.	A/A,G,H_2S	Glucose and lactose and/or fermentation, Gas produced, H_2S produced.
8	Red/Yellow with black precipitate.	K/A,H_2S	Glucose fermentation only, H_2S produced.
9	Yellow/Yellow with black precipitate.	A/A,H_2S	Glucose and lactose and/or fermentation, H_2S produced.

Clinical significance . .	The TSI medium facilitates preliminary identification of Gram negative bacilli.
Skills to be achieved . .	Skillfully perform the TSI test.
Skill evaluation criteria	1. Inoculation - 1mark. 2. Aseptic condition - 1 mark 3. Stabbing - 2 marks 4. Observing correctly : • Slant colour change - 2 marks. • Butt colour change - 2 marks. • Gas production - 2 marks. Total : 10 marks.
FAQs . .	What is the principle of TSI test ?
Assignment/Activity . .	Inoculate 4-5 TSI medium tubes.
Reference . .	Theory Topic 9 - Handbook of Microbiology

Week No.	. .	18th
Practical No.	. .	18
Title/Aim	. .	The student shall be able to do IMViC test - Indole test
Objectives	. .	The student shall be able to do Indole test
Principle	. .	Certain bacteria breakdown amino acid trptophan to give indole. Indole reacts with kovac's reagent to give red colour.
Requirements	. .	• Peptone water • Culture. • Kovac's reagent
Environment	. .	MLT laboratory
Procedure	. .	1. Subculture on media containing trptophan (e.g. peptone water) 2. Incubate at 37^{O}C For 24 hrs. 3. Add few drops of kovac's reagent 4. Examine for development of red ring.
Observations	. .	Positive test is identified by a development of red ring

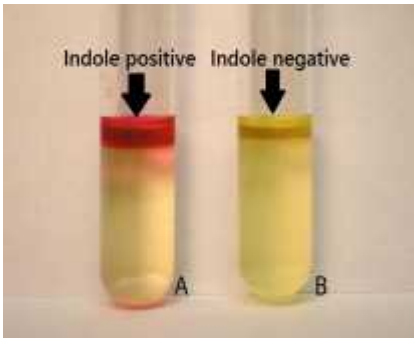


fig:18.1

Positive organisms : E.coli
Negative organism : Klebsiella

Result	. .	Development of red ring : Positive No development of red ring : Negative.
Clinical significance	. .	•The IMViC (indole) test will determine the biochemical properties and enzymatic reactions of enterobacteriaeceae. • The IMViC (indole) will determine whether they are pathogenic, occasional pathogenic or normal intestinal flora.
Skills to be achieved	. .	• Skillfully perform IMViC (indole) test under aseptic conditions with no contamination
Skill evaluating criteria		1. Inoculation into medium - 4 marks 2. Aseptic conditions - 4 marks 3. Differentiating between +ve and -ve test - 2 marks. Total : 10 marks.
FAQs		How you will do Indole test?
Assignment/Activity		Study Indole formation of 3-4 bacteria.
Reference		Theory Topic 9 - Handbook of Microbiology

Week No.	. .	19th week
Practical No.	. .	19
Title/ Aim	. .	To do IMViC - Methyl Red (MR) test
Objectives	. .	The student shall be able to do Methyl Red (MR) test
Principle	. .	This is to detect the ability of an organism to produce and maintain stable acid end products from glucose fermentation. Some bacteria produce large amounts of acids from glucose fermentation that they overcome the buffering action of the system. Methyl Red is a pH indicator, which remains red in color at a pH of 4.4 or less.
Requirements	. .	• Glucose phosphate broth • Methyl Red reagent • Culture • Inoculating loop • Incubator
Environment	. .	MLT laboratory
Procedure	. .	The bacterium to be tested is inoculated into glucose phosphate broth, which contains glucose and a phosphate buffer and incubated at 37^{o}C for 48 hours. Over the 48 hours the acid producing organism must produce sufficient acid to overcome the phosphate buffer and remain acid. The pH of the medium is tested by the addition of 5 drops of MR reagent. Development of red color is taken as positive. MR negative organism produce yellow colour.
Observations	. .	Development of red color is taken as positive. MR negative organism produce yellow colour

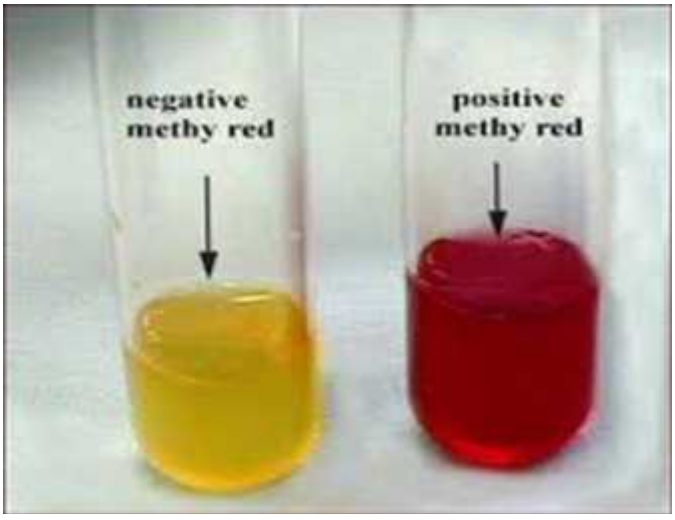


Fig:19.1

Eschericihia coli : Positive ; Klebsiella pneumoniae : Negative

Result . .	Development of red color is taken as positive. MR negative organism produce yellow colour
Clinical significance . .	The IMViC (MR) test will determine the biochemical properties and enzymatic reactions of enterobacteriaeceae. The IMViC (MR) will determine whether they are pathogenic, occasional pathogenic or normal intestinal flora.
Skillsto be achieved . .	Skillfully perform IMViC(MR) test under aseptic conditions with no contamination
Skill evaluating criteria	1. Inoculation into medium - 4 marks 2. Aseptic conditions - 4 marks 3. Differentiating between +ve and -ve test - 2 marks. Total : 10 marks.
FAQs . .	How you will do Methyl Red (MR) test?
Assignment/Activity . .	Perform Methyl Red test of 3-4 bacteria.
Reference . .	Theory Topic 9 - Handbook of Microbiology

Week No.	. .	20th week
Practical No.	. .	20
Title/Aim	. .	To do IMViC - Voges Proskauer (VP) Test
Objectives	. .	The student shall be able to doVoges Proskauer (VP) test
Principle	. .	VP test detects butylene glycol producers. Acetyl-methyl carbinol (acetoin) is an intermediate in the production of butylene glycol. If acetoin is present, it is oxidized in the presence of air and KOH to diacetyl. Diacetyl then reacts with guanidine components of peptone, in the presence of alpha-naphthol to produce red color.
Requirements	. .	• Glucose phosphate broth • Culture to be inoculated • Inoculating loop • Alpha-naphthol • 40% KOH
Environment	. .	MLT laboratory
Procedure	. .	1. Bacterium to be tested is inoculated into glucose phosphate broth and incubated for at least 48 hours. 2. 0.6 ml of alpha-naphthol is added to the test broth and shaken. 3. 0.2 ml of 40% KOH is added to the broth and shaken. 4. The tube is allowed to stand for 15 minutes. 5. Appearance of red color is taken as a positive test. 6. The negative tubes must be held for one hour, since maximum color development occurs within one hour after addition of reagents.

Observations

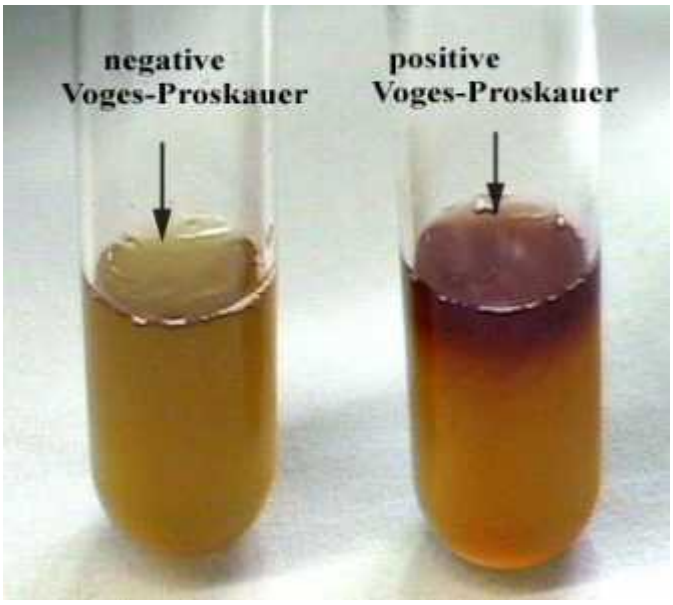


fig 20.1

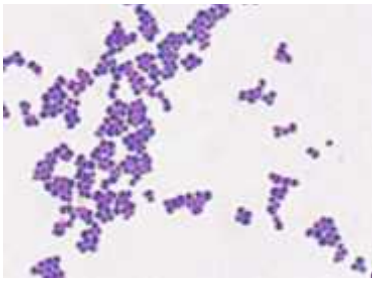
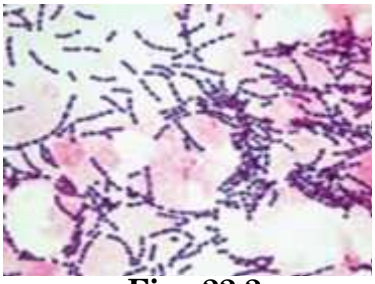
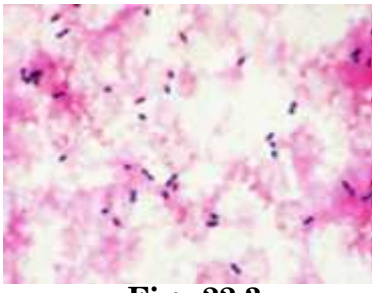
Escherichia coli : Negative ; Klebsiella pneumoniae : Positive

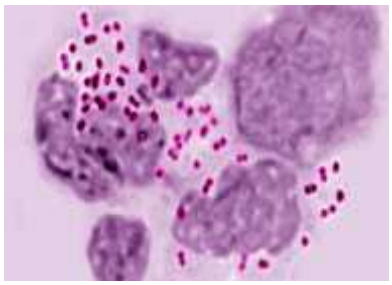
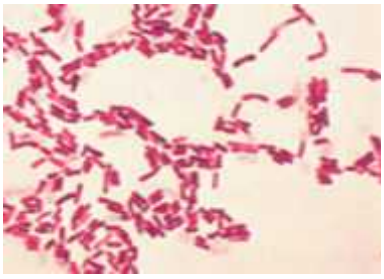
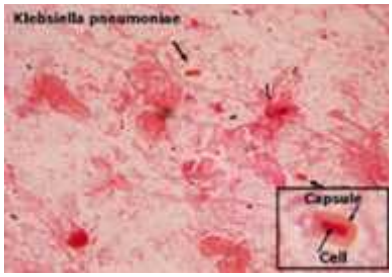
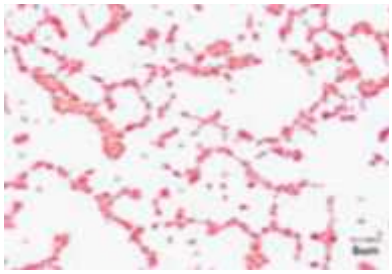
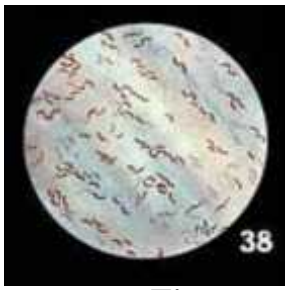
Result . .	Appearance of red color is taken as a positive test. The negative tubes must be held for one hour, since maximum color development occurs within one hour after addition of reagents.
Clinical significance . .	• The IMViC (VP) test will determine the biochemical properties and enzymatic reactions of enterobacteriaeceae. • The IMViC (VP) will determine whether they are pathogenic, occasional pathogenic or normal intestinal flora.
Skills to be achieved . .	Skillfully perform VP test under aseptic conditions with no contamination
Skill evaluating criteria	1. Inoculation in to medium - 4 marks 2. Aseptic conditions - 4 marks 3. Differentiating between +ve & –ve test - 2 marks Total : 10 marks.
FAQs . .	How you will do Voges Proskauer (VP) test?
Assignment/Activity . .	Do Voges Proskauer (VP) test of 3-4 bacteria.
Reference . .	Theory Topic 9 - Handbook of Microbiology

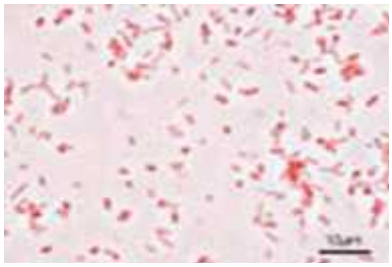
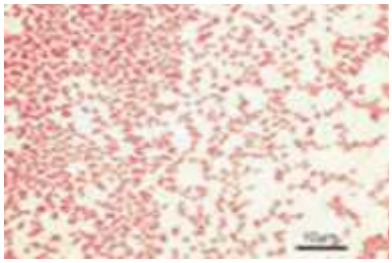
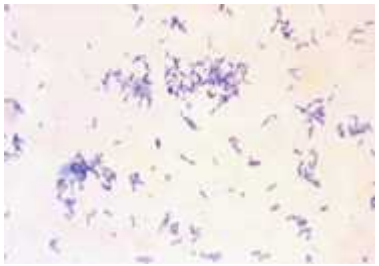
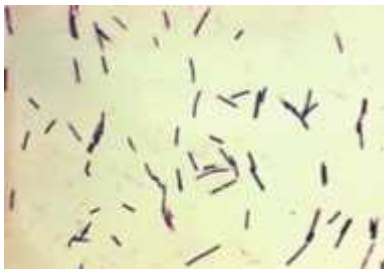
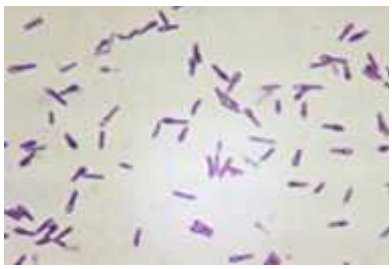
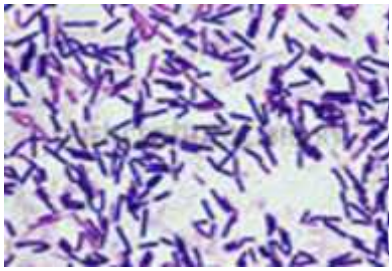
Week No. . .	21th week
Practical No. . .	21
Title/Aim . .	To do IMViC - Citrate Utilization test
Objectives . .	The student shall be able to do Citrate Utilization test.
Principle . .	This test detects the ability of an organism to utilize citrate as the sole source of carbon and energy. Bacteria are inoculated on a medium containing sodium citrate and a pH indicator bromothymol blue. Utilization of citrate involves the enzyme citritase, which breaks down citrate and results in alkaline pH. This results in change of medium's color from green to blue.
Requirements . .	• Simmon's citrate agar • Culture to be tested • Inoculating loop
Environment . .	MLT laboratory
Procedure . .	1. Bacterial colonies are picked up with a straight wire and inoculated into slope of Simmon's citrate agar and incubated overnight at 37°C. 2. If the organism has the ability to utilize citrate, the medium changes its color from green to blue.
Observations	 **Fig 21.1** Klebsiella pneumoniae: Positive Escherichia coli: Negative
Result . .	If the organism has the ability to utilize citrate, the medium changes its color from green to blue.
Clinical significance . .	• The IMViC (citrate test) will determine the biochemical properties and enzymatic reactions of enterobacteriaeceae. • The IMViC (citrate test) will determine whether they are pathogenic, occasional pathogenic or normal intestinal flora.
Skills to be achieved . .	Skillfully perform citrate test under aseptic conditions with no contamination
Skill evaluating criteria	1. Inoculation into medium - 4 marks 2. Aseptic conditions - 4 marks 3. Differentiating between +ve &–vetest - 2 marks Total : 10 marks.
FAQs . .	How you will do citrate utilization test?
Assignment/Activity . .	Study citrate utilization of 3-4 bacteria.
Reference . .	Theory Topic 9 - Handbook of Microbiology

Week No. . .	22st week
Practical No. . .	22
Title/ Aim . .	To identify the bacterium from permanent mounts
Objectives . .	To give knowledge to the student how to identify microorganisms on the basis of their gram staining characters and morphology.
Principle . .	Identification of bacterium.
Requirements . .	Permanent mounts of bacteria Microscope.
Environment . .	MLT laboratory
Procedure . .	Focus the given slide under oil immersion objective and report

Observations

Gram character	shape	Arrangement	Name of bacteria	Observation
Gram positive	Cocci	Grape-like clusters	Staphylococcus aureus	 **Fig: 22.1**
Gram positive	Cocci	Chains	Streptococcus pyogenes	 **Fig: 22.2**
Gram positive	Cocci	Lanceolate diplococcic	Pneumococci Streptococcus pneumoniae	 **Fig: 22.3**
Gram negative	Cocci	Diplococcic	Neisseria meningitides	 **Fig: 22.4**

Gram negative	Cocci	Diplococci kidney bean shaped.	Neisseria Gonorrhoeae	 **Fig: 22.5**
Gram negative	Bacilli	Single	Escherichia Coli	 **Fig: 22.6**
Gram negative	Bacilli	Single	Klebsiella pneumoniae	 **Fig: 22.7**
Gram negative	Bacilli	Single	Proteus	 **Fig: 22.8**
Gram negative	Bacilli	Comma shaped	Vibrio Cholerae	 **Fig: 22.9**
Gram negative	Bacilli	Single	Shigella	 **Fig: 22.10**

Gram negative	Bacilli	Single	Salmonella	 **Fig: 22.11**
Gram negative	Bacilli	Single	Pseudomonas	 **Fig: 22.12**
Gram positive	Bacilli	Chinese letter arrangement	Corynebacterium	 **Fig: 22.13**
Gram positive	Bacilli	Single with oval and subterminal spores	Clostridium welchii	 **Fig: 22.4**
Gram positive	Bacilli	Single with round and terminal spores	Clostridium tetani	 **Fig: 22.15**
Gram positive	Bacilli	Single with oval and subterminal	Clostridium botulinum	 **Fig: 22.16**

Result . .	Same as in table
Skills to be achieved . .	Knowledge of the identifying characteristics of given organism
Skill evaluating criteria	Each organism: a. Gram character - 2 marks b. Shape - 2 marks c. Arrangement - 2 marks d. Name- 2 marks e. Diagram - 2 marks. Total : 10 marks.
FAQs . .	State the gram character and morphology of given organism?
Assignment/Activity . .	Draw diagrams of given organisms focused under the microscope stating their gram character and morphology.
Reference	Theory topic :12 – Handbook of Microbiology

Week No.	. .	23nd week
Practical No.	. .	23
Title/Aim	. .	Bacteriological examination of Air, Water, Milk.
Objectives	. .	Student shall be able to perform bacteriological examination of Air, Water and Milk.
Requirements	. .	1. Water sample 2. Milk sample 3. Sterile water 4. Nutrient agar 5. Mac Conkey agar 6. Incubator
Environment	. .	MLT laboratory
Procedure		**Air Examination** • Place agar plate (containing selective or non-selective agar, depending on organism(s) of interest) on a flat surface in the test location, and remove the lid. • Leave the agar exposed for the agreed period of time (this may vary depending on the likely level of contamination in the test environment. But time periods of at least 30 minutes and up to 4 hours are usually recommended). Monitor the exposure time with a timer. • Replace the lid. Place the plates in an incubator overnight. • Count the number of colonies. • Perform biochemical tests to determine the organism. **Water examination** • The laboratory procedure involves making serial dilutions of the sample (1:10, 1:100, 1:1000, etc.) in sterile water and cultivating these on nutrient agar in a dish that is sealed and incubated. • MacConkey agar is used to count Gram-negative bacteria such as *E. coli*. • One set of plates is incubated at 22°C for 24 hours and a second set at 37°C for 24 hours. • The composition of the nutrient usually includes reagents that resist the growth of non-target organisms and make the target organism easily identified, often by a color change in the medium • At the end of the incubation period the colonies are counted by eye, a procedure that does not require a microscope as the colonies are typically a few mm **Milk examination** • Serial dilution of milk samples (1:10, 1:100, 1:1000, etc.) in sterile water is carried out to obtain the different dilutions. • These milk dilutions are further transferred into sterile nutrient agar Petri plates and distributed uniformly.

	• Nutrient agar plates are incubated for 24 hrs at 37°C. • Bacterial colonies are observed and counted after incubation and the count is multiplied by dilution factor.
Observations . .	Colonies on Nutrient Agar and Mac Conkey agar.
Result . .	The given air, water, milk sample contains bacteria.
Clinical significance . .	It is a method of analyzing air, water, milk to estimate the numbers of bacteria present and, if needed, to find out what sort of bacteria they are
Skills to be achieved . .	Air, Water, Milk analysis technique.
Skill evaluation criteria	1. Dilution technique - 4 marks 2. Plating technique - 4 marks 3. Observation - 2 marks. Total : 10 marks.
FAQs . .	How you will you do bacteriological examination of given air, water, milk sample?
Assignment/Activity . .	Analyse 4 to 5 air, water, milk sample for their bacterial content.
Reference . .	—

Week No.	. .	24th week
Practical No.	. .	24
Title/Aim	. .	To do Antibiotic Sensitivity Test (AST)
Objectives	. .	Student shall be able to perform Antibiotic Sensitivity Test.
Principle	. .	Whatman no. 2 filter paper circular disks (6 mm) impregnated with known concentration of antibiotics are placed on an Agar plate which is inoculated with a culture of the bacteria under test. The plate is incubated at 37°C for 24 hrs. During incubation, the antibiotics diffuse through the Agar. Susceptibility effectiveness is proportional to the diameter of the incubation zone around the disc. Organisms which row up to the age of the disc are resistant
Requirements	. .	• Nutrient broth • Muller Hinton Agar plates • Sterilized cotton swabs • Commercially available antibiotic discs
Environment	. .	MLT laboratory
Procedure	. .	1. Subculture bacteria from the isolated colonies in 1 ml of nutrient broth for 2-4 hrs. at 37°C. This subculture i used as inoculums for the antibiotic disc diffusion plate. 2. Divide the plate into sections according to the number of antibiotics. 3. Inoculate properly by using a sterile swab so as to obtain uniform distribution of the inoculums. 4. Place commercially available antibiotics discs on the inoculated plates using sterile forceps. 5. Incubate the plates overnight at 37°C 6. Measure the diameter of the zone of inhibition of growth in mm. 7. Report the result as follows:
Observations	. .	 **Figure 24.1**

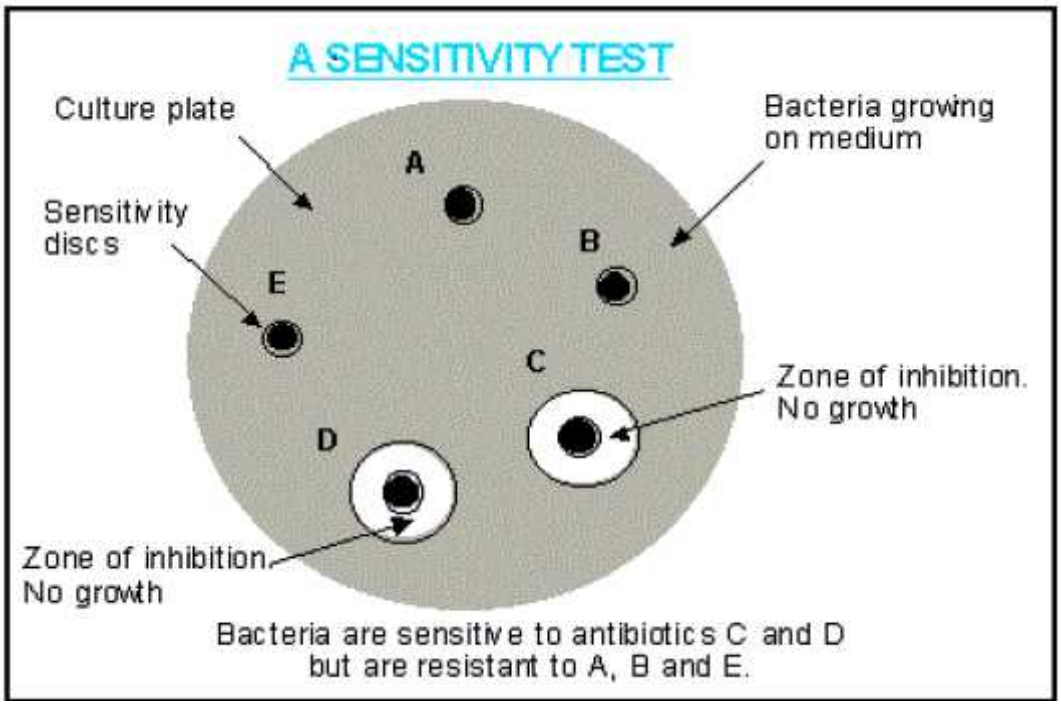
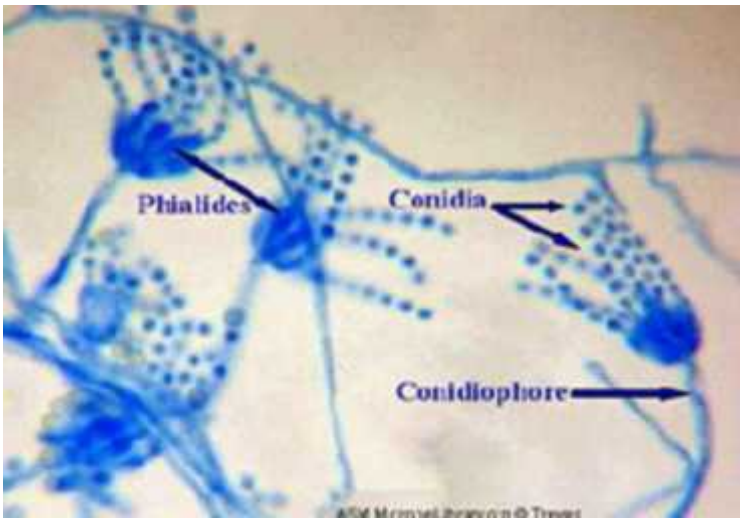


Figure 24.2 Antibiotic Sensitivity Test

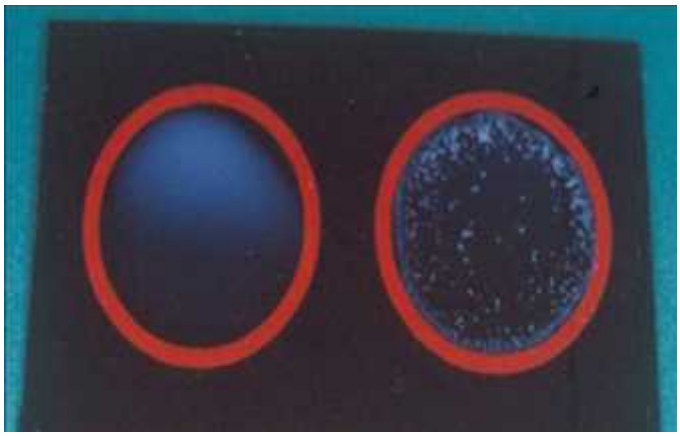
Result -

Observations	**Reports**
Zone less than 4 mm	Resistant
Zone 4-12 mm	Intermediate
Zone more than 12 mm	Sensitive

Clinical significance . .	Antibiotic sensitivity test is mainly useful if usually effective antibiotics fail to produce the desired effect. In the treatment and control of infectious diseases which are caused by pathogens that are drug resistant, sensitivity testing is helpful in selecting effective antimicrobial drugs. These tests measure antimicrobial activity against bacteria under laboratory conditions and not in the patient.
Skills to be achieved . .	Skillfully perform AST
Skill evaluation criteria	1. Plating technique - 2 marks 2. Aseptic condition - 2 marks 3. Selection of disc - 1 mark. 4. Placing the disc on the agar plate - 2 marks 5. Zone measurement - 2 marks. 6. Sensitivity reporting - 1 mark Total : 10 marks.
FAQ . .	How you will perform AST for given organism?
Assignment/Activity . .	Do AST of 3-4 organisms.
Reference . .	Ref. Theory topic17 - Handbook of Microbiology

Week No. . .	25th week
Practical No. . .	25
Title/ Aim . .	Fungal examination by wet mount.
Objectives . .	Students shall be able to stain the fungal preparation by lacto phenol cotton blue
Principle . .	Lactic acid preserves the fungal structure and clears the tissue, phenol acts as a disinfectant.
Requirements . .	• Lacto phenol cotton blue • Fungal specimen
Environment . .	MLT laboratory
Procedure . .	1. Place a drop of lacto phenol cotton blue on a clean slide. 2. Place fungal specimen on the drop and tease gently. 3. Place a coverslip on the mount and press down gently. 4. Examine under low power first and then under high power
Observations	
	 **Figure 25.1 Aspergillus, Stained with Lacto Phenol Cotton Blue**
Result . .	Fungal material appears pale to dark blue.
Clinical significance . .	Fungal infection.
Skills to be achieved . .	Skillfully prepare wet mount of fungal specimen and observe under the microscope.
Skill evaluation criteria	1. Preparing wet mount with no air bubbles- 4 marks. 2. Proper teasing - 2 marks. 3. Focusing - 2 marks. 4. Identification - 2 marks. Total : 10 marks.
FAQs . .	How you will prepare wet mount of given fungal specimen
Assignment/Activity . .	Prepare wet mount of 4-5 fungal specimen
Reference . .	—

Week No. . .	26th week
Practical No. . .	26
Title/ Aim	Serology
	RA test (Rheumatoid Arthritis test)
Objectives . .	The student shall be able to perform R.A. test accurately.
Principle . .	Small latex particles are coated with specially treated IgG. When serum containing Rheumatoid Factor is mixed with the IgG-coated latex particles, the rheumatoid factors gets bound to the IgG and cause agglutination.
Requirements . .	1. Serum sample. 2. Kit contains :— a. RF Antigen b. Positive control c. Negative control d. Glass slide with reaction circles e. Rubber teats.
Environment . .	MLT laboratory
Procedure . .	**Qualitative methods** 1. Pipette one drop of serum on to the glass slide using the disposable pipette provided with the kit. 2. Add one drop of RHELAX RF latex reagents to the drops of serum on the slide. Do not let the dropper tip touch the liquid on slide. 3. Using a mixing sticks mix the serum and the RHELAX RF factor reagent uniformly over entire circles. 4. Immediately start a stop watch. Rock the slide gently back and forth, observing for agglutination macroscopically at 2 mins. **Semi Quantitative methods :** 1. Using isotonic solution prepare serials dilutions of the serum sample positive in the quantitative method 1:2, 1:4, 1:8, 1:32, 1:64, and so on. 2. Pipette each dilution of serum on to the separate reaction circles. 3. Add one drop of RHELAX RF latex reagents to each drop of a diluted serum sample on the slide. Do not let the dropper tip touch the liquid on the slide. 4. Using a mixing sticks, mix the sample and the latex reagents uniformly over the entire circles. 5. Immediately start a stopwatch. Rock the slide gently back and forth, observing for agglutination macroscopically at 2 mins.

Observations	 **Figure 26.1 RA test**
Result . .	**Interpretation of Test Result :** **Qualitative method :** • Agglutination is a positive test result and indicates the pleasance of rheumatoid factors in the test specimen. • No agglutination is a negative tests result and indicates the absence rheumatoid factors in the test specimen.
Clinical significance . .	Helps in the detection of Rheumatoid arthritis.
Skills to be achieved . .	Skillfully perform RA test
Skill evaluation criteria	1. Dilution technique - 4 marks 2. Addition of correct reagents in proper sequence- 3 marks. 3. Addition of correct samples to correct circle- 2 marks. 4. Observation - 1 mark. Total : 10 marks.
FAQs . .	1. How you will do qualitative RA test ? 2. How you will do quantitative RA test ?
Assignment/Activity . .	Perform RA test of 3 - 4 samples.
Reference . .	Theory Chapter 20 - Handbook of Microbiology

Week No.	. .	27th week
Practical No.	. .	27
Title/ Aim	. .	To do VDRL test
Objectives	. .	Student shall be able to perform VDRL test in the laboratory.
Principle	. .	Complement present in serum interferes with the Flocculation reaction. It is inactivated by keeping the serum at 56°C. Antibodies of Syphilis react with the VDRL antigen (particles of lipid coated with cardiolipin) and forms floccules.
Requirements	. .	1. Fasting serum specimen 2. VDRL plate 3. Commercially available VDRL kit containing • VDRL Carbon Antigen Suspension, • Positive Control Serum, • Negative Control Serum, • Test Cards, • Mixing Sticks. • Buffered saline • Normal Saline Note : Prepare a working VDRL antigen fresh before the test. Follow manufacturer's instructions.
Environment	. .	MLT laboratory
Procedure	. .	**Method 1 : Slide test** **Procedure--** • Keep 0.5 ml of serum in a water bath at 56°C for inactivation for 30 minutes. • Cool to room temperature. • Add about 0.05 ml inactivated serum in the cavity of VDRL plate. • Add one drop of working VDRL antigen to the cavity containing serum. • Rotate the plate on the rotor immediately for four minutes or by hand ten times in five seconds. • Examine visually in bright light and confirm by observing under low power objective. • Process one negative and one positive control serum in same manner.

Method 2 : Tube test

Procedure—

- Label test tubes 1 to 9.
- Prepare serial dilution of inactivated serum.
  - o Pipette o.1 ml of serum in tube no.1.
  - o Add 0.1 ml of normal saline and mix (dilution 1:2)
  - o Transfer 0.1 ml of dilution 1:2 to tube no. 2 add 0.1 ml of saline and mix (dilution 1:4)
  - o Similarly prepare other dilutions (i.e. 1:8, 1:16, 1:32, 1:64, 1:128, 1:256)
- Add 0.5 ml of each dilution to the numbered cavities of VDRL slide.
- Add one drop of freshly prepared VDRL antigen to the diluted serum on the VDRL slide.
- Rotate the plate for 4 min.
- Observe the floccules

Observations

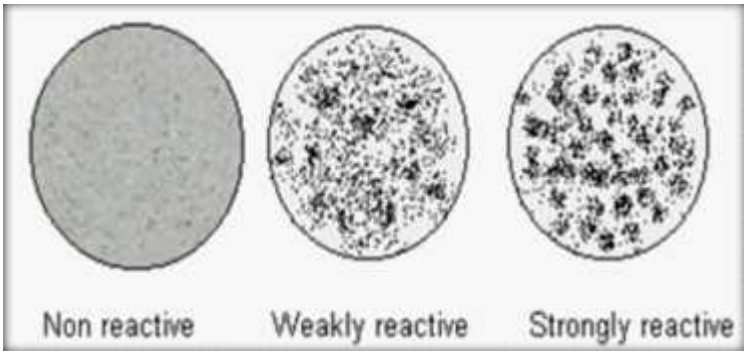


fig:27.1

Result—

- Report as follows:

Test Report	Observation
VDRL test Non-Reactive . .	No clumps
VDRL test Weakly Reactive . .	Small clumps with free particles.
VDRL test Reactive . .	Medium and large clumps on a clear background

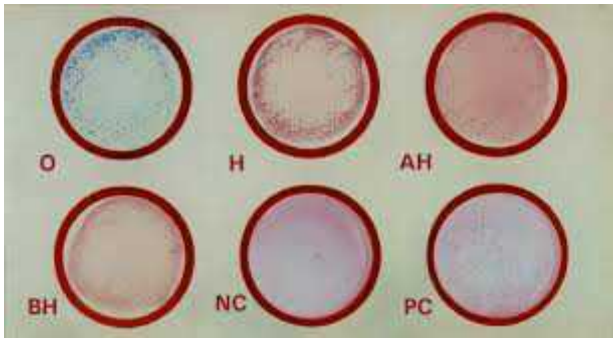
Observations

Tube method

- Report as follows :

Tube no.	1	2	3	4	5	6	7	8
Dilution	1:2	1:4	1:8	1:16	1:32	1:64	1:128	1:256
Reaction	R	R	R	R	R	WR	NR	NR

KEY : **R=Reactive, WR= weakly reactive, NR= nonreactive** Note : In case of weakly reactive serum, use dilution up to 1:32.	
Clinical significance . .	VDRL test includes determination of serological response to Treponema infection Syphilis
Skills to be achieved . .	Skillfully perform VDRL test
Skill evaluation . . **criteria** . .	1. Dilution technique- 4 marks 2. Addition of correct reagents in proper sequence- 3 marks. 3. Addition of correct samples to correct circle- 2 marks. 4. Observation- 1 mark. Total : 10 marks.
FAQs . .	How you will do VDRL test Qualitative and Quantitative ?
Assignment/Activity . .	Do VDRL test of 5 samples
Reference . .	Ref. Theory chapter 20- Handbook of Microbiology

Week No. . .	28th week
Practical No. . .	28
Title/ Aim . .	To do Widal test by Slide method
Objectives . .	The student must be able to perform Widal test accurately and independently using Slide method.
Principle . .	Antibodies found in patients serum in response to exposure to Salmonella organisms agglutinate a bacterial suspension of Salmonella (non infective), which carries homologous antigens.
Requirements . .	1. Specimen : Serum (fresh) 2. Commercial kits containing four specific antigen suspensions :— • Salmonella typhi 'O' • Salmonella typhi 'H' • Salmonella paratyphi 'AH' • Salmonella paratyphi 'BH' 3. Slides with ceramic rings. 4. Droppers 5. Applicator sticks.
Environment . .	MLT laboratory
Procedure . .	**Slide method** 1. Mark the circles on the glass plate as O, H, AH, BH. 2. Add one drop of serum to each of the circles. 3. Add corresponding antigen one drop to the marked circles. 4. Mix antigen and serum of each circle using separate applicator sticks 5. Slowly rock and tilt the glass plate for 3mins and observe for agglutination. **Note :** A slight change in procedure may exist depending on the kit manufactures instructions.
Observations . .	**figure 28.1 Widal test - slide method** 

Result . .	Agglutination seen/not seen in each circle.
Clinical significance . .	Diagnosis of typhoid and paratyphoid
Skills achieved	Performing Widal test by slide method accurately
Skill evaluation criteria	1. Dilution technique - 4 marks 2. Addition of correct reagents in proper sequence- 3 marks. 3. Addition of correct samples to correct circle- 2 marks. 4. Observation - 1 mark. Total : 10 marks.
FAQs . .	1. Do Widal test by slide method of given specimen and report 2. Name the antigens which you will use for the test ?
Assignment/Activity . .	Do Widal test by slide method of given 4 - 5 specimen.
Reference . .	Theory chapter 20 – Handbook of Microbiology.

Week No. . .	29th week
Practical No.	29
Title/ Aim . .	To do Widal test by tube method.
Objectives . .	The student must be able to perform Widal test accurately and independently using tube method.
Principle . .	Antibodies found in patients serum in response to exposure to Salmonella organisms agglutinate a bacterial suspension of Salmonella (non infective), which carries homologous antigens.
Requirements . .	1. Specimen: Serum (fresh) 2. Commercial kits containing • 5 test tubes • 36 Widal tubes 3. Widal rack 4. Serological pipettes 0.1ml and 1.0 ml. 5. Serological water bath / incubator 6. Normal saline (NS)
Environment . .	MLT laboratory
Procedure . .	**Procedure :** 1. Collect 5 ml of blood by venous puncture in a plane tube. Allow it to clot. Separate the serum. 2. Make 1 in 10 dilution of patient's serum in 0.85% Normal saline (NaCl). 3. Place 8 small tubes in a row in a rack. 4. Like this arrange four rows for salmonella typhi 'O' H' 'AH' and 'BH'. 5. Take 0.5 ml of normal saline in test tube No. 2 to 8. 6. Add 0.5 ml of patient's diluted serum in tube No. 1 and 2. 7. Mix and withdraw 0.5 ml from tube 2 into tube 3. 8. Mix and withdraw 0.5 ml from tube 3 into tube 4. 9. Continue till tube no. 7. 10. Withdraw and discard 0.5 ml from tube 7. 11. Tube No. 8 contains only normal saline. 12. Carry same procedure for all four rows. 13. Add 0.5ml of 'O', 'H', 'AH' and 'BH' in all tubes of respective rows. 14. Shake the rack and keep in an incubator at 37°C. 15. Observe for agglutination. 16. Find out the highest dilution which shows agglutination.

Tube no.	1	2	3	4	5	6	7	8
1st row (TH Antigen) 0.85% saline	Nil	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml
Patients diluted serum 1:10	0.5ml	0.5ml	0.5ml of tube no. 2	0.5ml of tube no. 3	0.5ml of tube no. 4	0.5ml of tube no. 5	0.5ml of tube no. 6 and discard 0.5ml from tube no. 7	Nil
TH antigen	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml	0.5ml control. No serum
Final dilution	1/20	1/40	1/80	1/160	1/320	1/640	1/1280	——-

Prepare similarly for other antigens i.e. O, AH, BH.

Observations

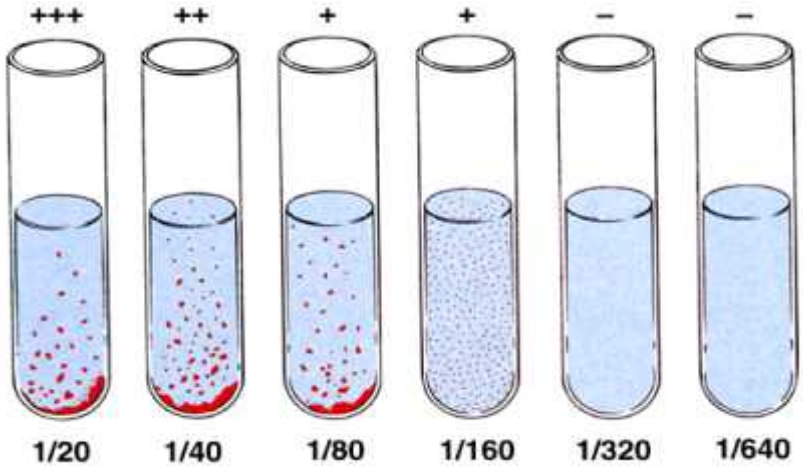


Fig. 29.1 Widal test (tube method)

Report :

Dilution	Antigen 'O'	Antigen 'H'	Antigen 'AH'	Antigen 'BH'
1:40				
1:80				
1:160				
1:320				
1:640				

Positive result should be correlated with clinical findings and precious history immunization.

Clinical significance . .	Diagnosis of typhoid and paratyphoid
Skills achieved . .	Performing Widal test by tube method accurately
Skill evaluation criteria	1. Dilution technique- 4 marks 2. Addition of correct reagents in proper sequence- 3 marks. 3. Addition of correct samples to correct circle-2 marks. 4. Observation - 1 mark. Total : 10 marks.
FAQs . .	1. Do Widal test by tube method of given specimen and report. 2. Name the antigens which you will use for the test?
Assignment/Activity . .	Do Widal test by slide method of given 4 - 5 specimen
Reference . .	Theory Chapter 20 - Handbook of Microbiology

(G.C.P.) Ra 755—9 (1500—9–2015)

Week No.	. .	30th week
Practical No.	. .	30
Title/ Aim	. .	ELISA Test (Enzyme-linked Immunosorbent Assay)
Objectives	. .	The student shall be able to do ELISA test for detection of HIV antibody
Principle	. .	HIV antigen is fixed on the surface of a test well. Patients serum (if anti HIV is present) will attach to the HIV antigen, after washing, an enzyme conjugate reagent is added and re-incubated. The enzyme conjugate antihuman IgG attaches to the HIV antigen-antibody complex. After washing, a substrate – chromogen reagent is added. This is acted on the enzyme and a colour is produced. A stop reagent stops the reaction. The color is read spectrophotometrically
Requirements	. .	1. Patients serum 2. Spectrophotometer 3. HIV – ELISA Kit containing • Control serum • Diluents buffer • Enzyme conjugates, etc. • Substrate – chromogen • Stop solution • Wash solution
Environment	. .	MLT laboratory
Procedure	. .	1. Add indicated amount of diluent of the micro plate well. 2. Add serum sample and controls to the separate wells (amount as specified in kit procedure) 3. Shake and incubate at 37°C for 1 hr. 4. Wash with wash solution. 5. Add enzyme conjugate, and incubate at 37°C for say 30 mins. 6. Wash with wash solution. 7. Add substrate – chromogen reagent. 8. Incubate at 37°C for say 20 mins. 9. Stop reagents is added to stop the reaction. 10. Color produced in test and control wells is measured spectrophotometrically.

Observations

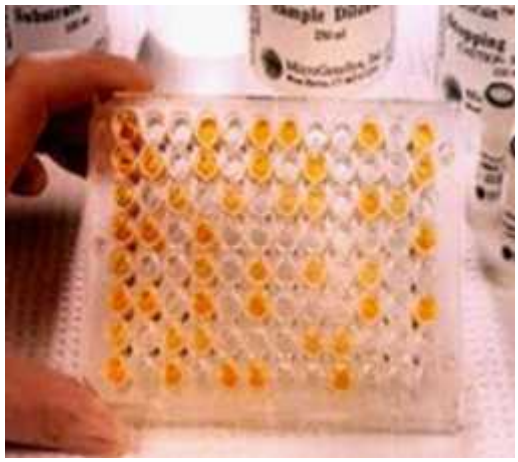
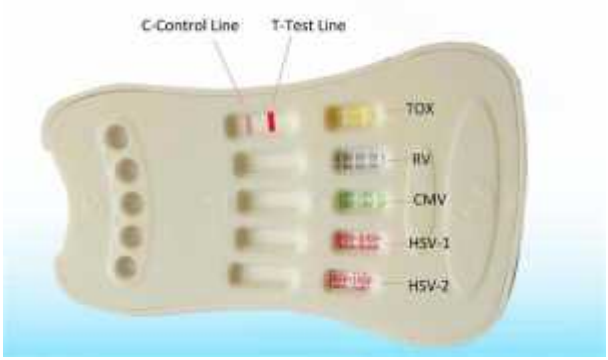


Fig:30.1 **ELISA Test**

Result . .	The given specimen is positive/ negative for ELISA test.
Clinical significance . .	ELISA test for detection of HIV antibody.
Skills to be achieved . .	Performing HIV test by ELISA method accurately.
Skill evaluation criteria	1. Stepwise addition of correct reagents specimen to the respective test wells - 6 marks. 2. Observing colour produced - 2 marks. 3. Difference between +ve & –ve test - 2 marks. Total : 10 marks.
FAQs . .	1. Do HIV test by ELISA method of given specimen and report 2. Name the reagents stepwise which you will use for the test?
Assignment/Activity . .	Do HIV test by ELISA method of given 4 - 5 specimen.
Reference . .	Theory Chapter 20 - Handbook of Microbiology

Week No.	. .	31st week
Practical No.	. .	31
Title/ Aim	. .	TORCH ELISA Test
Objectives	. .	The student shall be able to know about the TORCH ELISA test
Principle	. .	Same as ELISA method provided in the kit
Requirements	. .	Sample to be tested, Reagents supplied in the kit
Environment	. .	MLT laboratory/hospital visit.
Procedure	. .	**1. Toxoplasma gondii (toxoplasmosis)** A parasite that can be acquired from ingesting cysts from the feces of infected cats, drinking unpasteurized milk, or eating undercooked contaminated meat. Infection early in pregnancy can cause miscarriage. Later in pregnancy it can cause eye infections, and mental retardation. The detection of IgG/ IgM specific antibodies to T.gondii is particularly helpful for diagnosis of acute or primary infections. The determination is done by using diagnostic kit of ELISA. **2. Rubella (German Measles)** Infection early in pregnancy can cause birth defects such as heart disease, growth retardation and eye defect. The absence of Rubella specific IgG antibodies in sera of long term duration after primary infections in presence of virus specific IgM antibodies is indicative of risk of defects in newborn infants. This test provides the clinician a help for monitoring of the immunological response upon vaccination. The test is carried out using diagnostic kit of ELISA for Rubella virus. **3. Cytomegalo virus (CMV)** This virus is transmitted through body secretions (including breast milk) as well as sexual contact. Infection can cause death, hearing loss and mental retardation. The detection of virus specific IgM antibodies is of great value in the diagnosis of acute primary virus infections in the absence of typical clinical symptoms. Asymptomatic infections usually happen for Cytomegalo virus in healthy individuals during pregnancy. The test is carried out using diagnostic kit of ELISA for Cytomegalo virus. **4. Herpes simplex virus (HSV)** It is a common infection that spreads by oral and genital contact. The detection of virus specific IgM antibodies is of great value in the diagnosis of acute primary virus infections in the absence of typical clinical symptoms. Asymptomatic infections usually happen for Herpes II in healthy individuals during pregnancy. The test provides the clinician a help for monitoring of risk population and for the follow-up of primary infections. The test is carried out using diagnostic kit of ELISA for Herpes II.

Observations	 **Figure 31.1** **TORCH Test Kit**
Result . .	The given specimen is positive/negative
Clinical significance . .	ELISA test performed for TORCH panel are mainly to detect the presence or absence of specific antibodies in serum for following organisms: A. Toxoplasma gondii (TO) B. Rubella (R) C. Cytomegalo virus (C) D. Herpes II (H) TORCH infections are very common in adult population. Pregnant woman are exposed to these infections. Fatal infection usually occurs by the trans placental root after invasion of the mother's bloodstream
Skills to be achieved . .	Performing TORCH test by ELISA method accurately
Skill evaluation criteria	1. Knowledge of different tests - 4 marks. 2. Use of test - 4 marks. 3. Observation of tests - 2 marks. Total : 10 marks.
FAQs . .	How you will do TORCH test by ELISA method of given specimen?
Assignment/Activity . .	Prepare a PPT of procedure manual of different manufacturer's kits available in market.
Reference . .	Theory Chapter 20 - Handbook of Microbiology

Week No.	. .	32nd week
Practical No.	. .	32
Title/Aim	. .	Immunological Pregnancy Test
Objectives	. .	The student shall be able to do pregnancy test accurately.
Principle	. .	Monoclonal antibodies to HCG have been produced. These have been attached to enzymes which can later interact with a dye molecule and produce a color change.
Requirements	. .	Pregnancy test kit 1. Pregnancy test kit contains three regions – Reaction region, Test region and Control region • Reaction region: Contains monoclonal anti-HCG antibodies linked to enzyme. • Test region: contains polyclonal anti-HCG antibodies, which will bind to HCG molecules bound to monoclonal anti-HCG antibodies. Also contains dye molecules which will be activated if monoclonal antibodies bind to polyclonal anti-HCG antibodies. • Control region: Contains anti-mouse antibodies and dye molecules which will be activated if monoclonal antibodies bind here.
Environment	. .	MLT laboratory
Procedure	. .	1. When pregnant woman's urine travels up the pregnancy test, pregrancy test kit, HCG will bind tomonoclonal antibodies in reaction region. 2. Movement of the urine will move the monoclonal antibodies up to the test region 3. Monoclonal antibodies with bound HCG will bind to antibodies in test region and activate dye molecules, producing a color change 4. Any unbound monoclonal antibodies will continue to travel to control region and will bind to anti-mouse antibodies, activating dye molecules and producing a color change.
Observations		

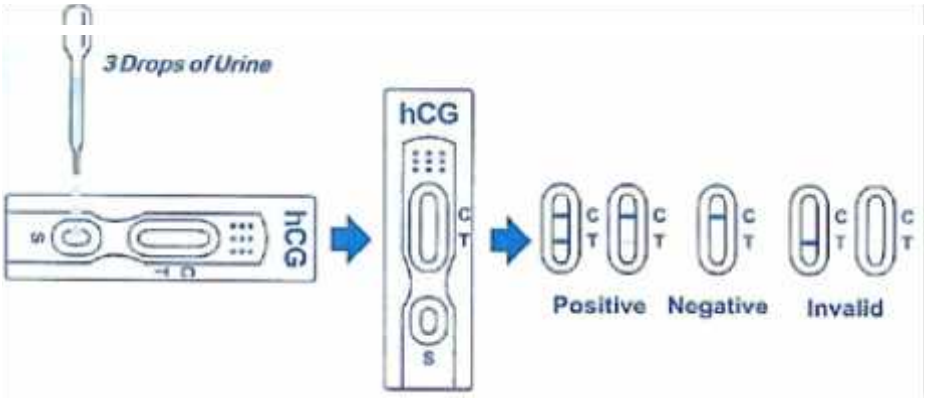


Figure32.1 Pregnancy test Results

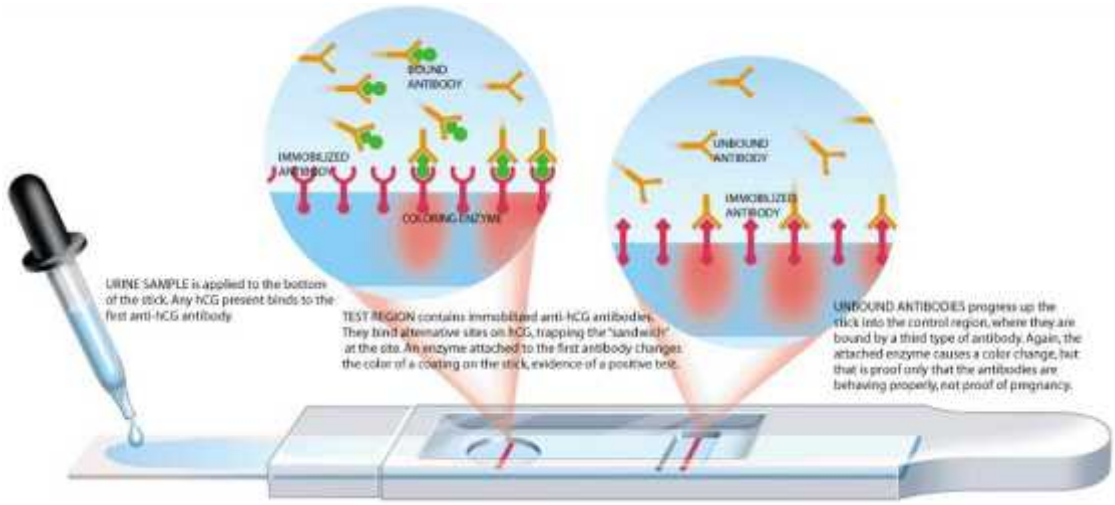


Figure 32.2 Pregnancy Tests – Mechanism

Result . .	The given urine specimen is positive/negative for pregnancy test.
Clinical significance . .	A pregnant woman has the hormone Human Chorionic Gonadotrophin (HCG) in her urine which is tested and accordingly pregnancy report submitted.
Skills to be achieved . .	Performing pregnancy test of urine sample accurately.
Skill evaluation criteria	1. Patient instructions - 2 marks 2. Addition of correct reagents - 4 marks 3. Sample addition - 2 marks 4. Correct reporting - 2 marks. Total : 10 marks.
FAQs . .	Do pregnancy test of urine sample and report
Assignment/Activity . .	Do pregnancy test of urine sample (4 - 5 specimen) and report
Reference . .	Theory Chapter 20 - Handbook of Microbiology

Week No.	. .	33rd Week
Practical No.	. .	33
Title/Aim	. .	**C-reactive protein test**
Objectives	. .	Student shall be able to perform C-reactive protein test in the laboratory.
Principle	. .	Latex particles coated with anti- CRP, agglutinate in the presence of CRP.
Requirements	. .	• Latex CRP kit • Normal saline • Serological pipettes 0.1ml and 1.0 ml.
Environment	. .	MLT laboratory
Procedure	. .	1. Dilute patient serum 1:5 with normal saline. 2. Place 1 drop of dil. Serum on the test slide. 3. Place 1 drop of positive and negative control also in the respective zone of the test slide. 4. Add one drop of reagent to each zone. 5. Mix well and observe for agglutinations.
Result	. .	Marked agglutinations generally indicate presence of CRP concentration above 6 mg/l
Clinical significance	. .	The C-reactive protein is normal alpha globulin and it is elevated in patients who have an inflammatory condition of infectious or non-infectious origin. The results are simple index of the disease activity and treatment status.
Skills to be achieved	. .	Performing C-reactive protein test in lab.
Skill evaluation criteria		• Proper dilution - 4 marks. • Use of test - 4 marks. • Observation of tests - 2 marks. Total : 10 marks.
FAQs	. .	1. Perform C-reactive protein test of given specimen and report 2. Name the reagents stepwise which you will use for the test?
Assignment/Activity	. .	Perform C-reactive protein test of given 4 -5 specimen.
Reference	. .	Theory topic -Handbook of Microbiology

Serum dilution	CRP mg/l
1:10	12
1:20	24
1:30	48
1:40	96